



*Interaction of *E. coli* with the intestinal immune system of the pig in the post-weaning period*



Eric Cox, Laboratory of Immunology, Fac. Vet. Med., UGent, Belgium
Eric.Cox@Ugent.be

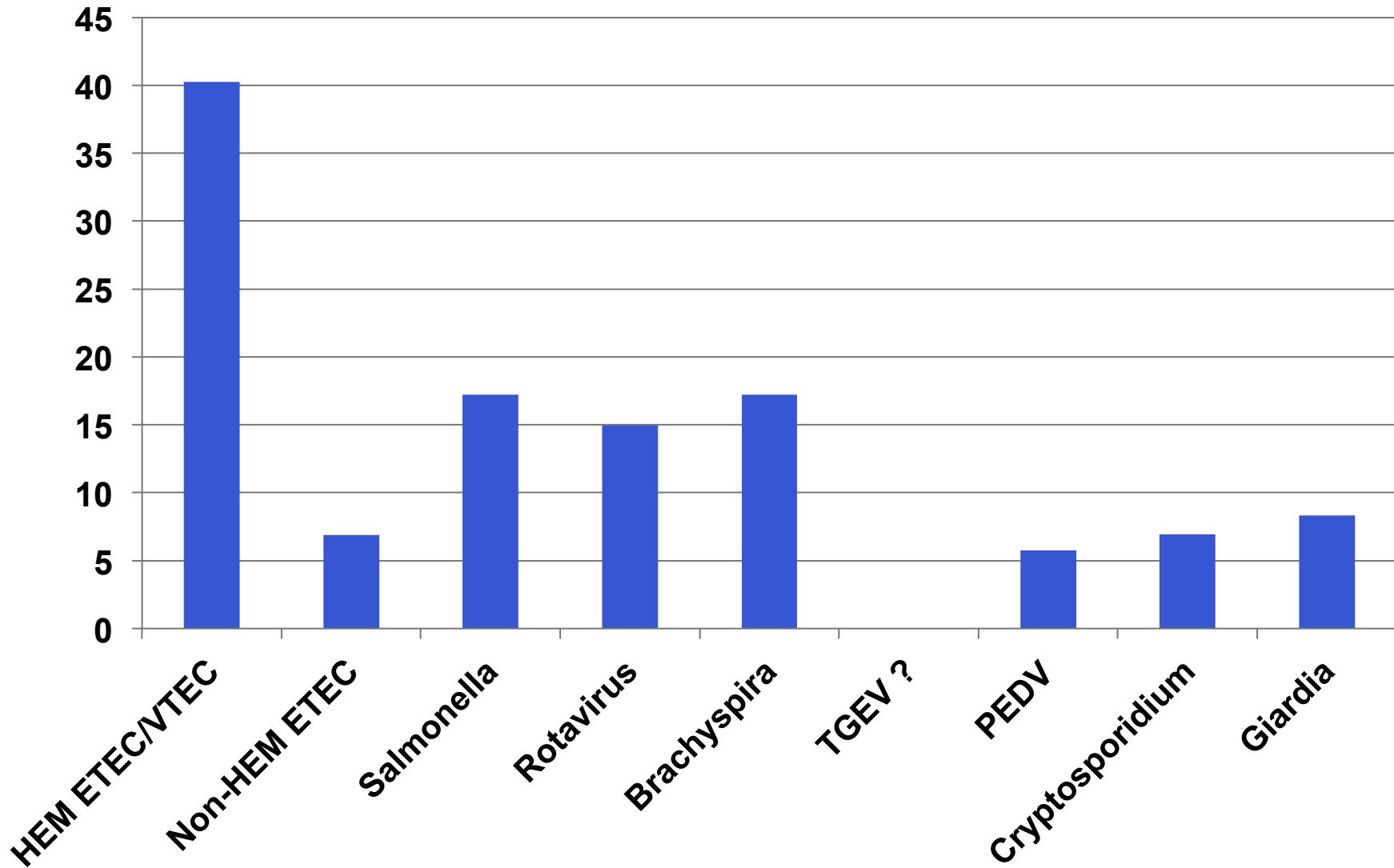
Pigs

- **Diarrhoea = 11 % of all post-weaning mortality**
- **± 10 million piglets die annually world-wide**
- **50 % is caused by enterotoxigenic *E. coli***
- **The economical losses due to oedema disease are not known**

Bacteria and viruses identified in faeces of pigs post weaning on Belgian farms

Coddens et al., manuscript in preparation

Percentage enteropathogens



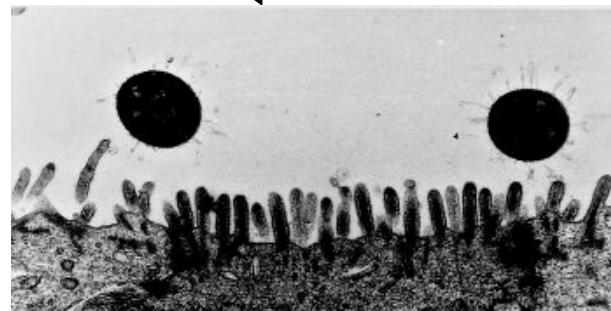
Piglets post-weaning



Enterotoxigenic *E. coli* (ETEC)
Vérotoxinogenic *E. coli* (VTEC)

Fimbriae (F4 (K88), F18)

Colonization factors
bind to sugars



Enterotoxins (LT, STa, STb, EAST1)

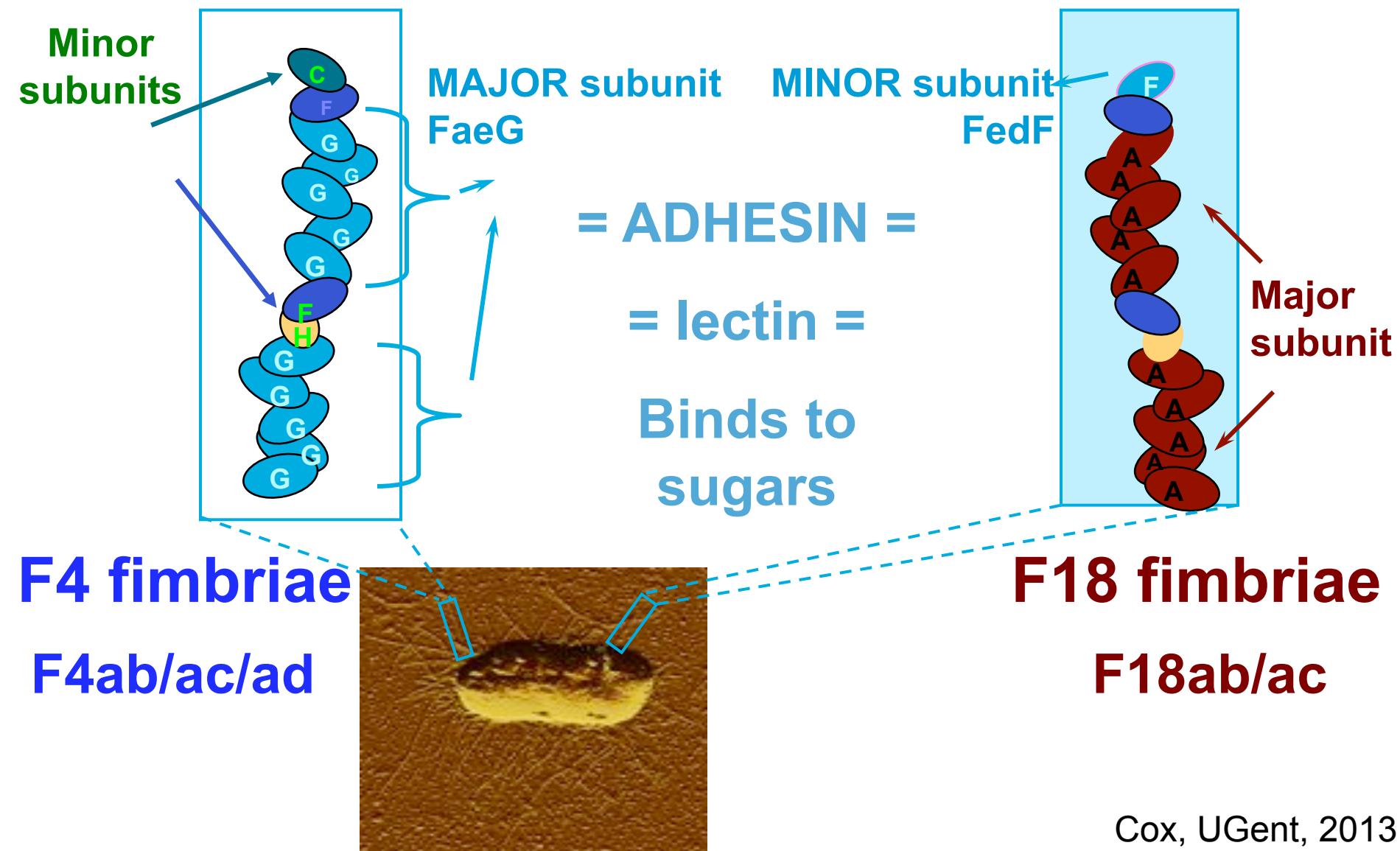
Shiga-toxin (Stx2e)

Post-weaning
diarrhoea

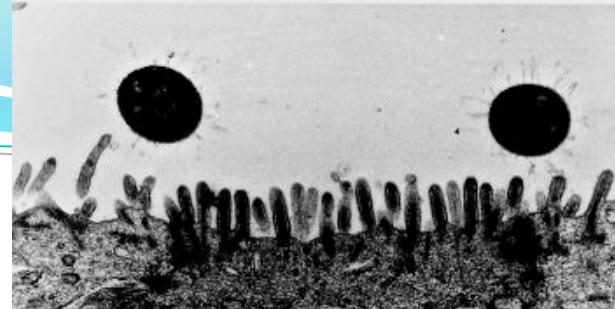
Oedema disease

The colonisation factors F4 and F18 and their F18 receptors

F4 and F18 fimbriae differ in structure and in receptor-specificity



F4 receptor phenotype



Phenotype	Adhesiveness	Receptor	Identification of receptor	
			Characterization	Molecular mass (kDa)
A	ab, ac, ad	<i>bcd</i>	glycoproteins	45–70
		<i>bc</i>	glycoproteins	210 and 240
B	ab, ac	<i>bc</i>	glycoproteins	210 and 240
C	ab, ad	<i>d</i>	glycosphingolipid	?
D	ad	<i>d</i>	glycosphingolipid	?
E	/	/	Receptor negative phenotype	
F	ab	<i>b</i>	glycoprotein	74

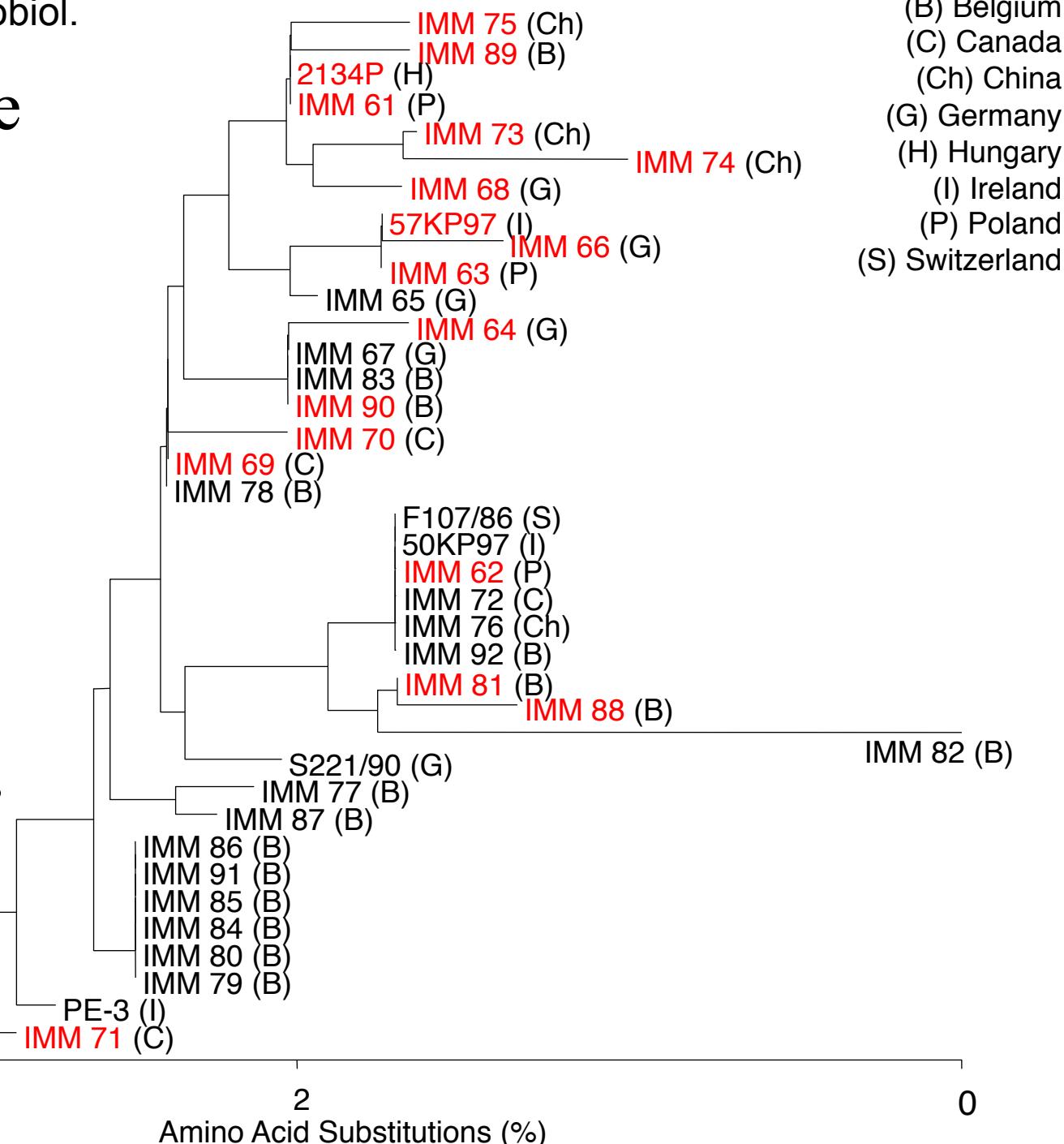
F4 binding occurs to the sugars and is complex

Phylogenetic tree for FedF the adhesine of F18

Differences in FedF
between F18ab &
F18ac?

Region specific
variations?

Each strain has
maximal 11 variations
96 %



Conclusions

F4ab ≠ F4ac ≠ F4ad

FedF 96% identical

No difference between F18ab⁺ & F18ac⁺ *E. coli*

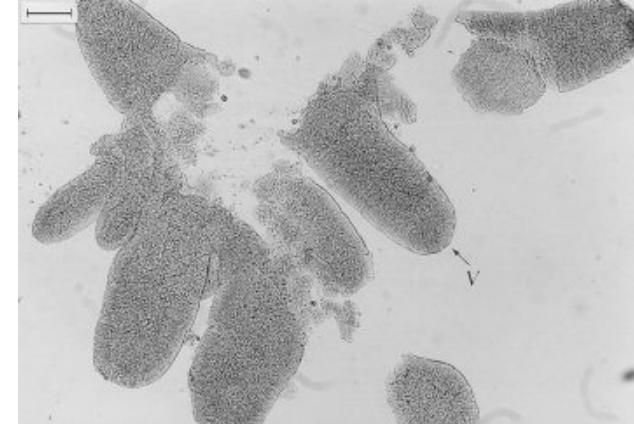
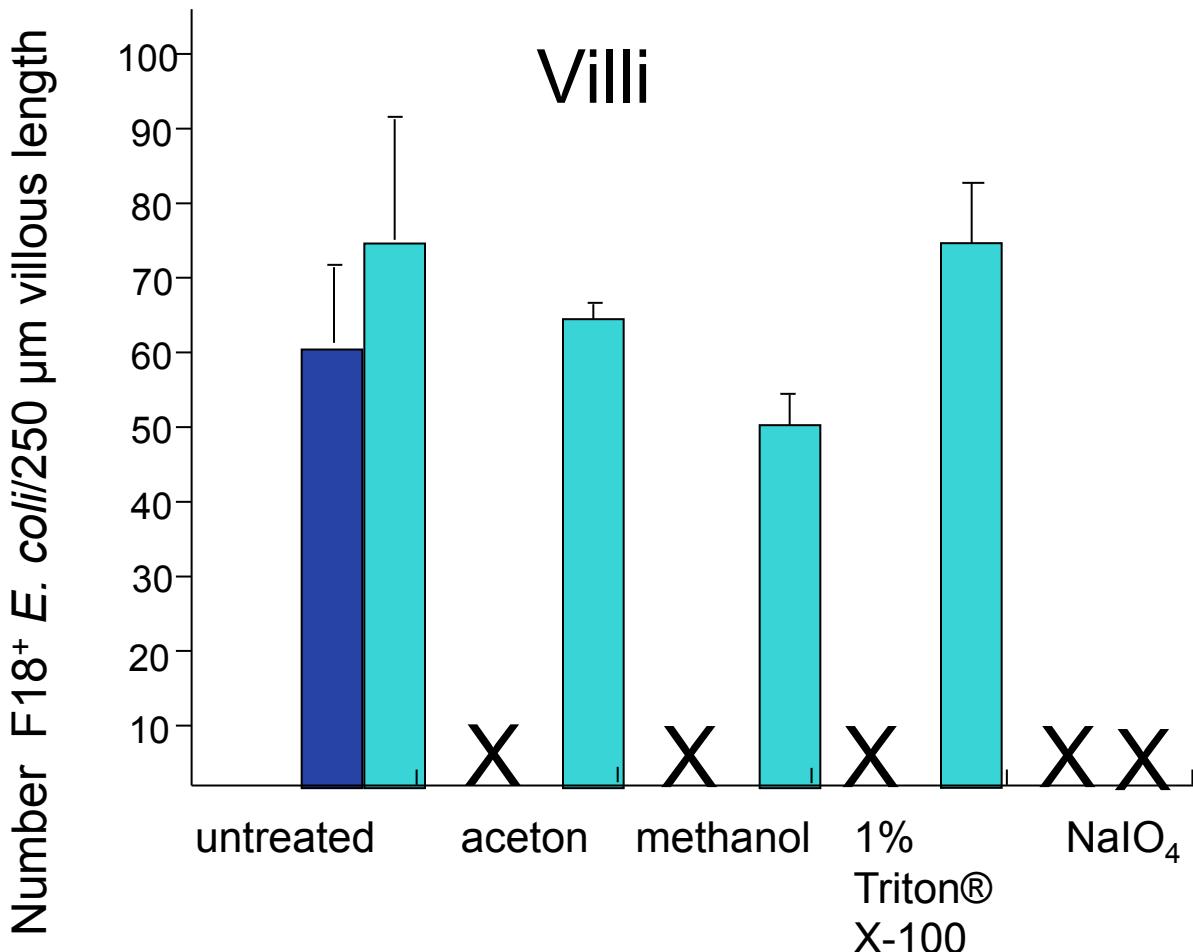
No region specific variation

FedF is worldwide conserved

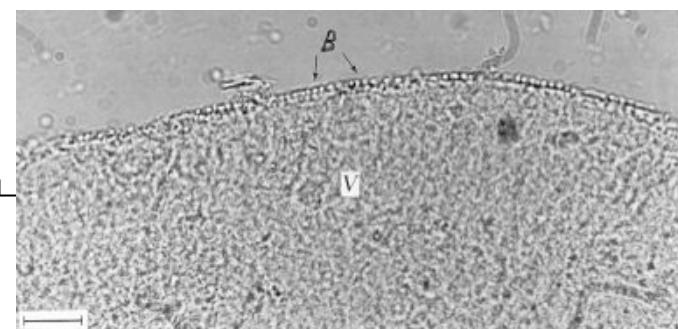


The same receptors

Wat is the nature of the F18 receptor ?



F18⁺ E. coli (n = 4)
or
F4ac⁺ E. coli (n = 2)



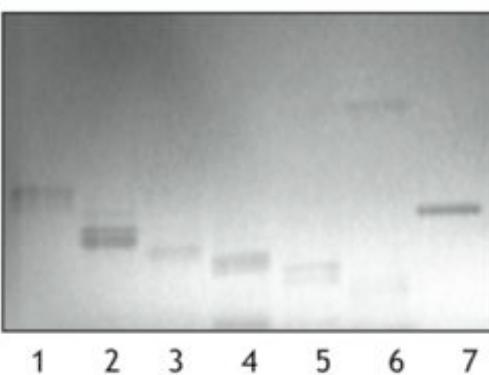
⇒ The F18R is a glycolipid

⇒ The F4R is a glycoprotein

Coddens et al., 2009. J. Biol. Chem.

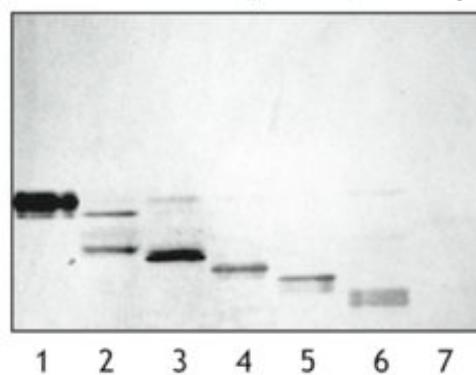
Isolation and characterization of F18⁺ *E. coli*-binding glycosphingolipids from blood group O and A pig intestinal epithelium

A. Chemical detection



1 2 3 4 5 6 7

B. *E. coli* HB101(pIH120) + FedF



1 2 3 4 5 6 7
↑ ↑ ↑ ↑ ↑ ↑

Non-acid glycosphingolipid fraction

Mass spectrometry

Proton NMR spectroscopy

Blood group O =>

H5 type 1 Fucalpha2Galbeta3GlcNAcbeta3Galbeta4Glcbeta1Cer

Blood group A =>

A6 type 1 GalNAca3(Fucalpha2)Galbeta3GlcNAcbeta3Galbeta4Glcbeta1Cer

A7 type 4 GalNAca3(Fucalpha2)Galbeta3GalNAcbeta3Galalpha4Galbeta4Glcbeta1Cer

A8 type 1 GalNAca3(Fucalpha2)Galbeta3GlcNAcbeta3Galbeta3GlcNAcbeta3Galbeta4Glcbeta1Cer

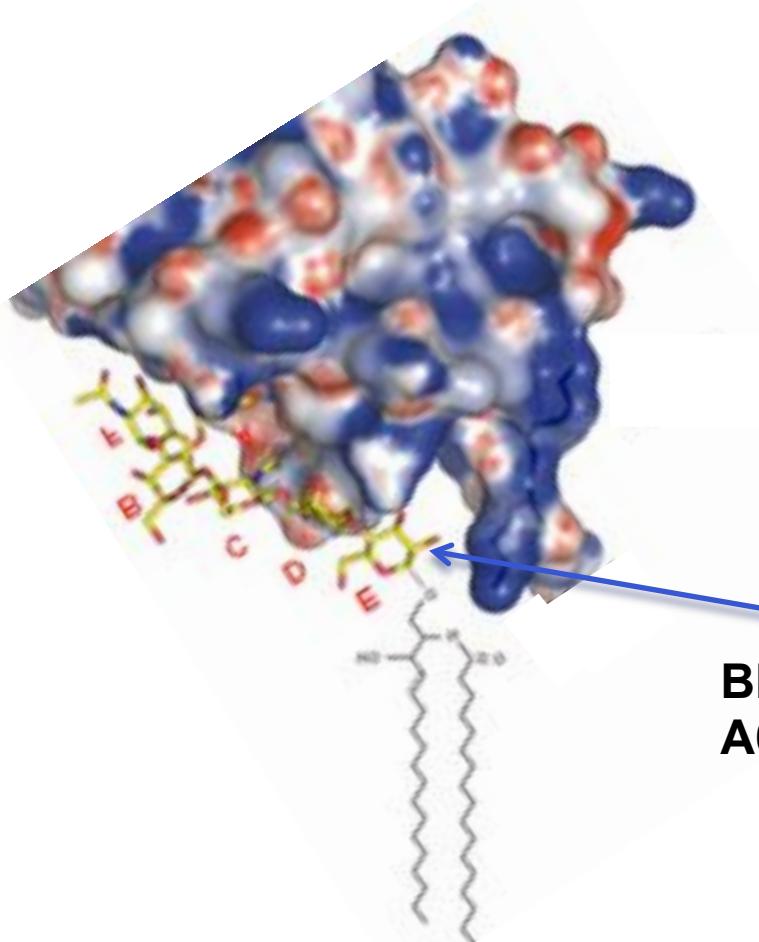
A9 type 1 GalNAca3(Fucalpha2)Galbeta3GalNAca3(Fucalpha2)Galbeta3GlcNAcbeta3Galbeta4Glcbeta1Cer

Decaglycosylceramide with terminal HexNAc-(Fuc-)Hex-HexNAc sequence

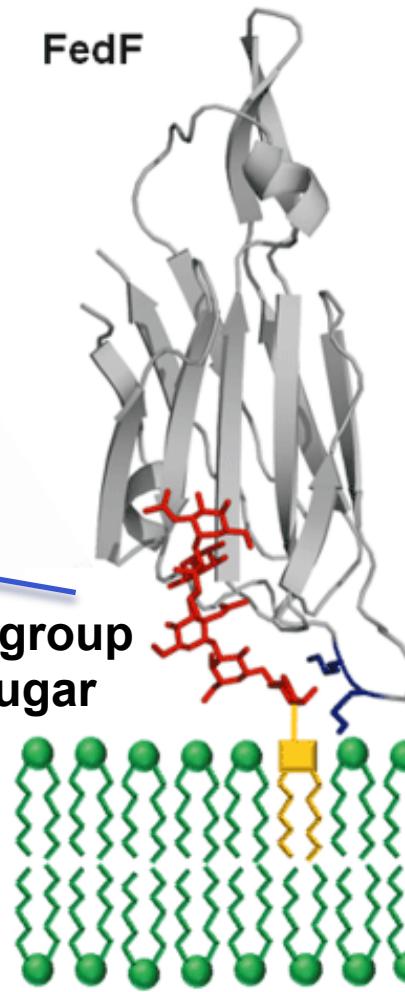
Undecaglycosylceramide with terminal HexNAc-(Fuc-)Hex-HexNAc sequence

Dodecaglycosylceramide with terminal HexNAc-(Fuc-)Hex-HexNAc sequence

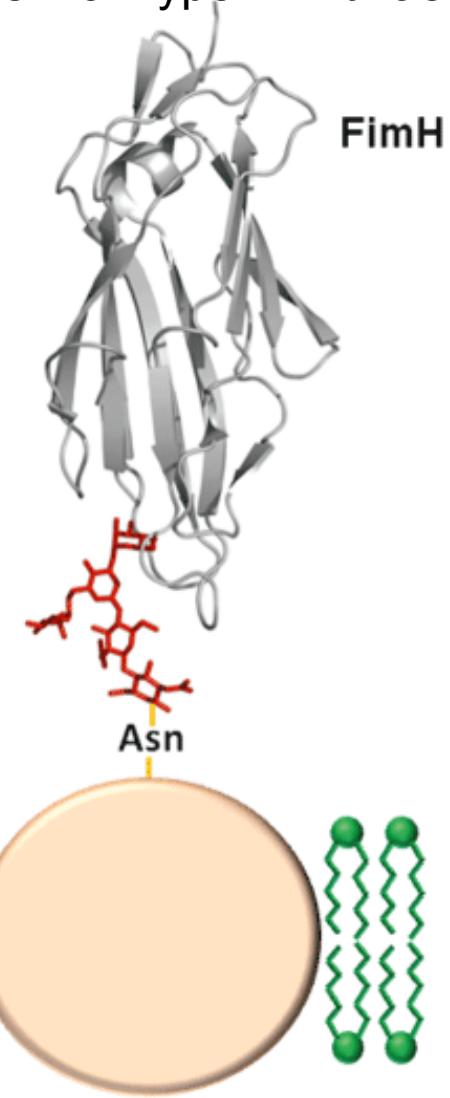
Structural insight in histo-blood group binding by the F18 fimbrial adhesin FedF



Adhesin of F18 fimbriae
FedF



Adhesin of Type I fimbriae

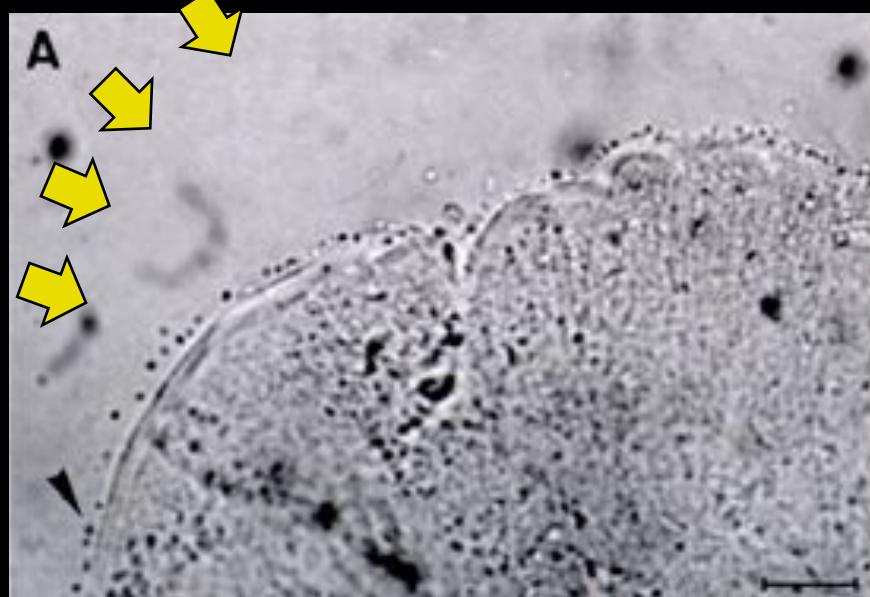


Conclusion

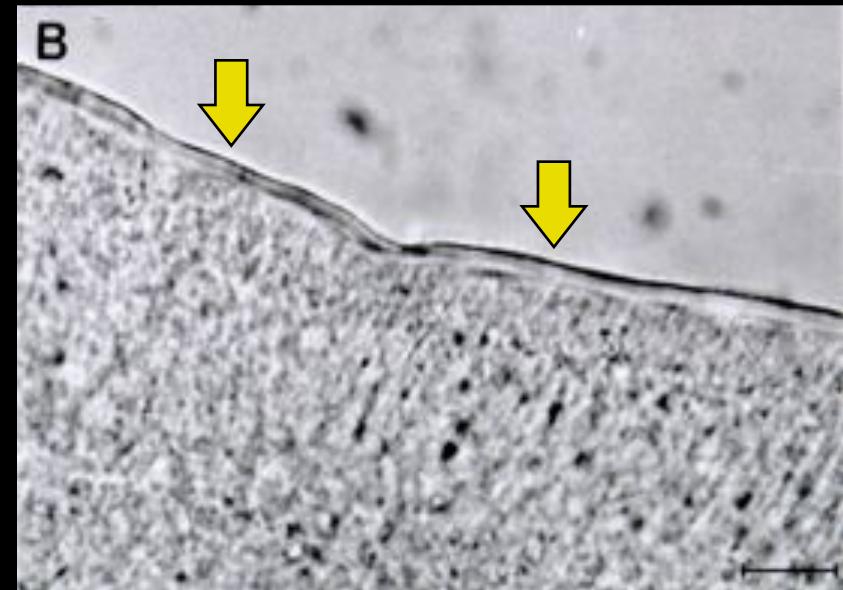
**F18 adheres to blood group A (A6,
A8, A9 type 1, A7 type 4) and O (H5
type 1) sugars**

The expression of fimbrial receptors as determined by *in vitro* adhesion

F4R +

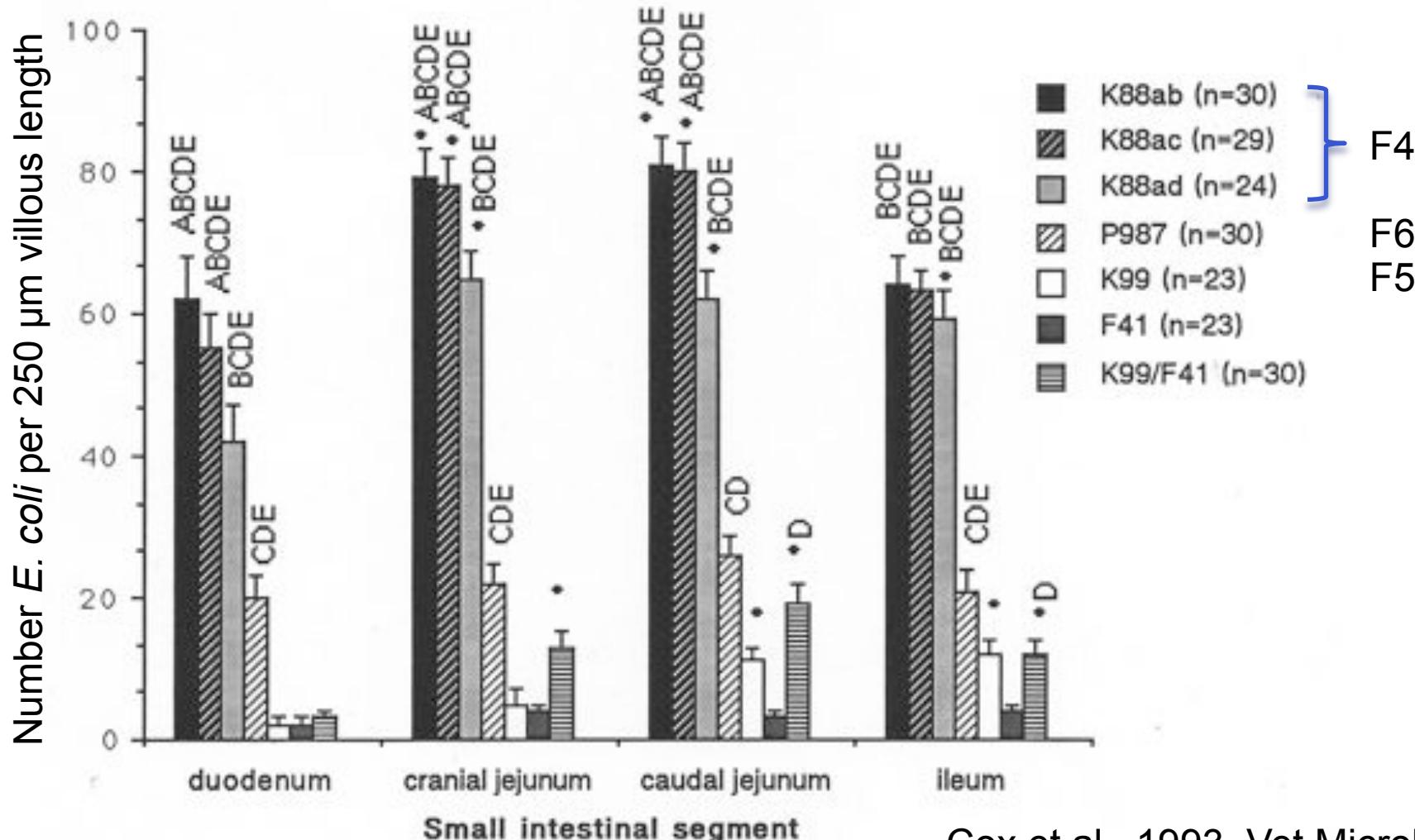
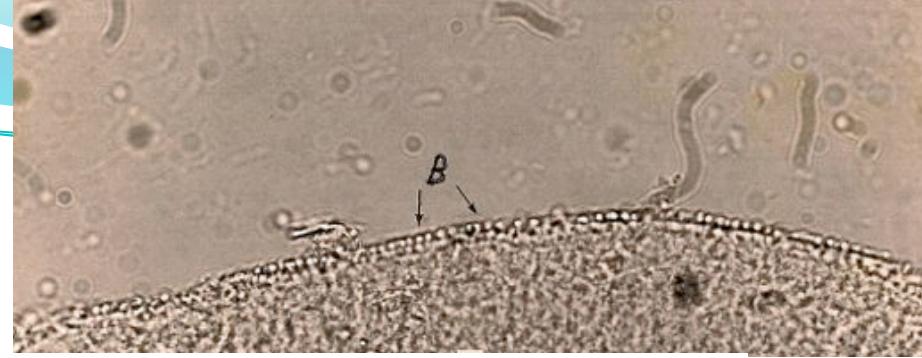


F4R- (Natural
knock-out)

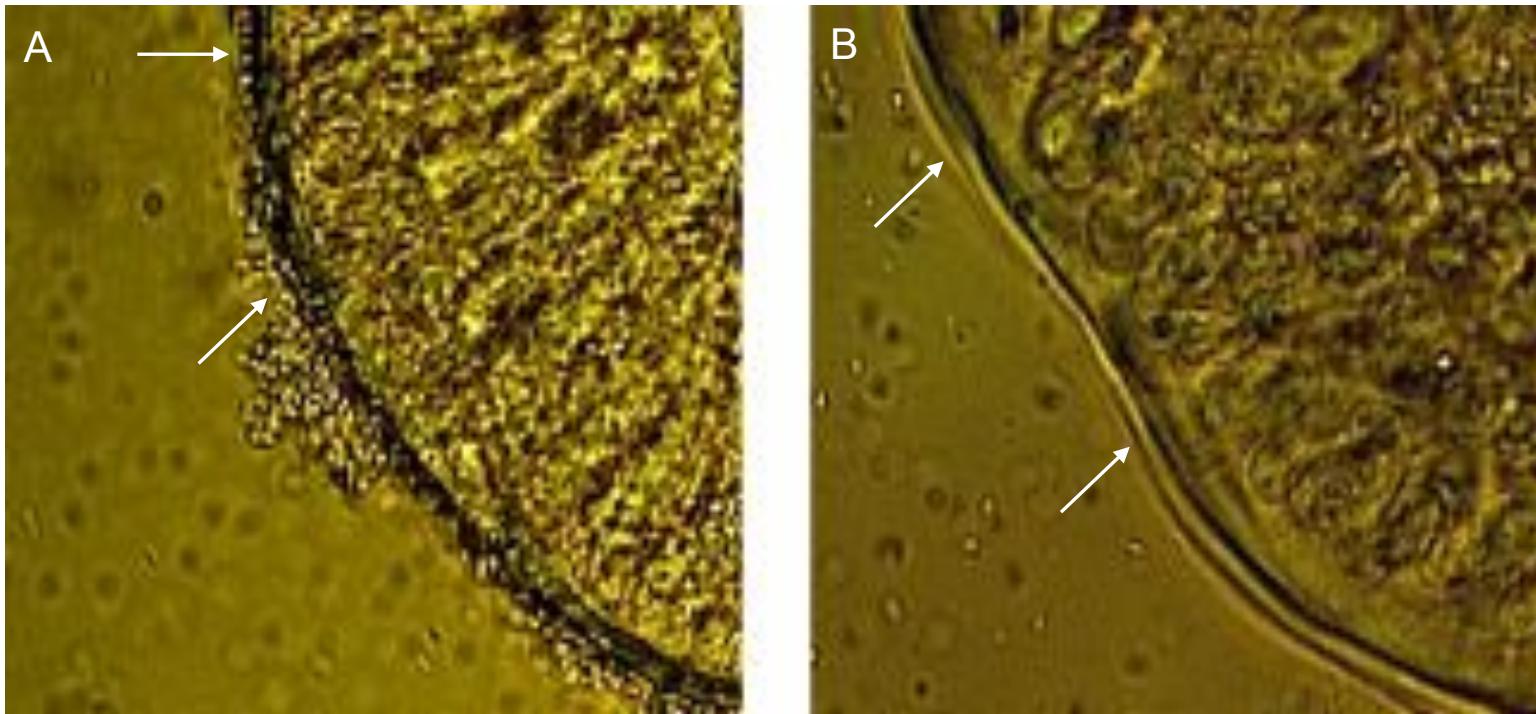


bar: 10 μ m

Binding of ETEC to villi of 4- to 5-week-old piglets



F18 receptors



In vitro villous adhesion assay

F18⁺ *E. coli* adhesion



F18R positive piglets

No adhesion



F18R negative piglets

Expression of F18R is function of age

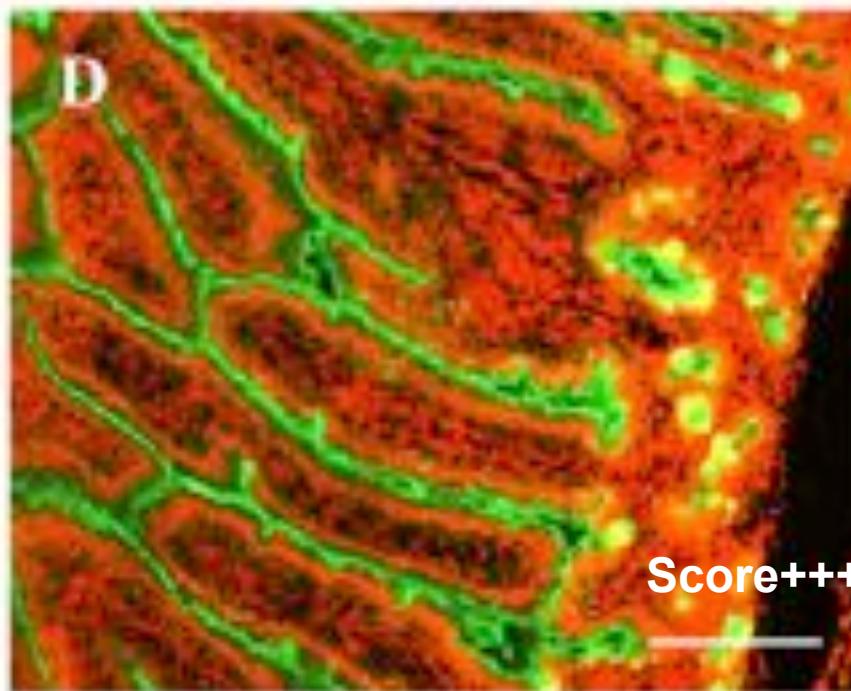
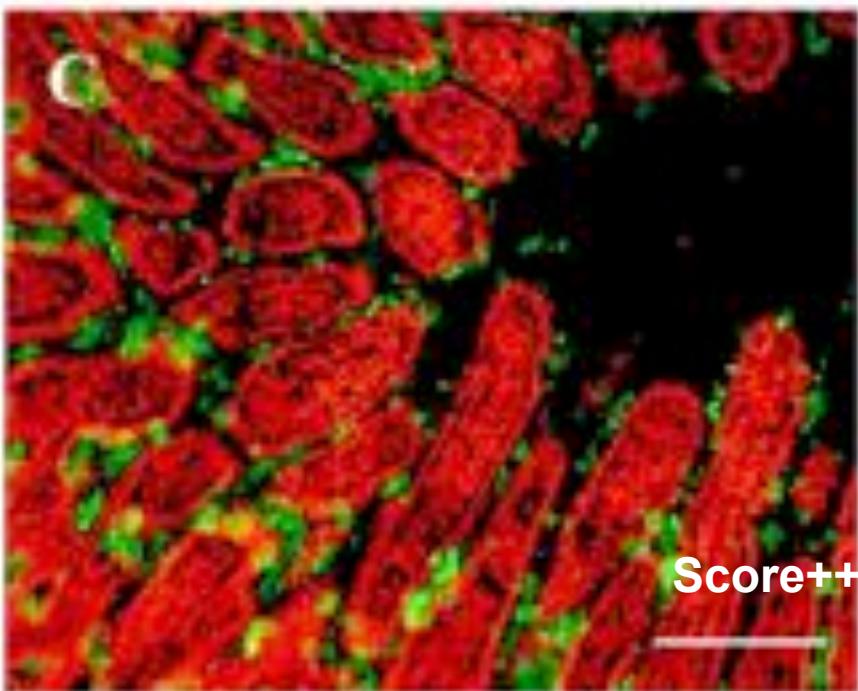
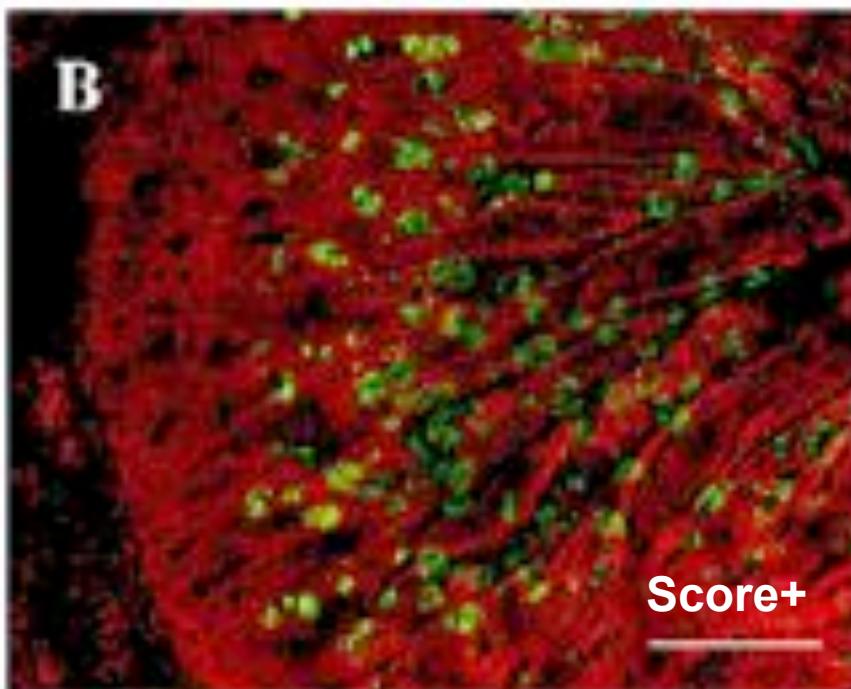
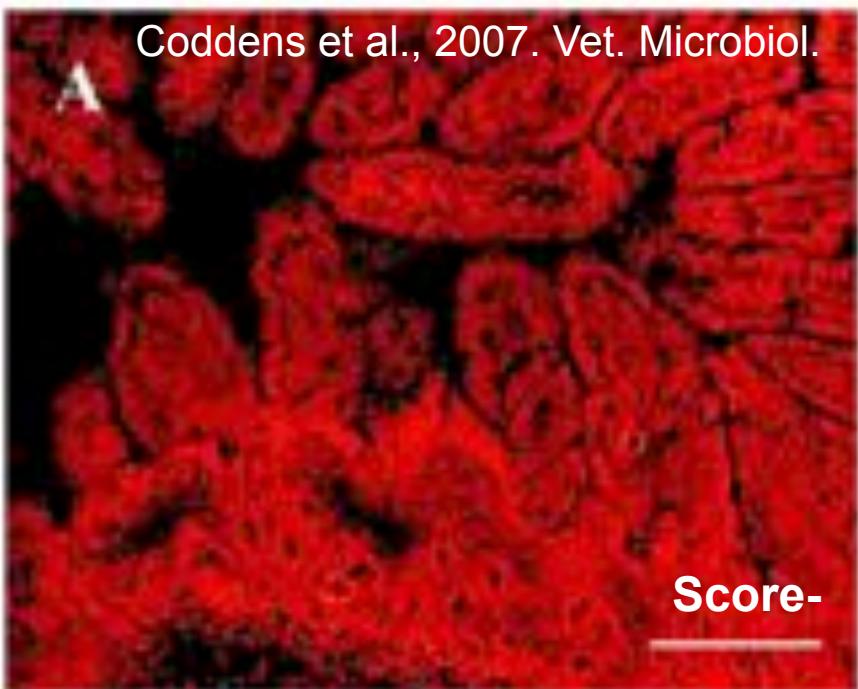
74 porcelets were tested

Number of piglets	Age (weeks)	FUT Expression			Number of piglets with F18+ <i>E. coli</i> adhering to 250 µm villous length		
		G/G Susceptible	G/A	A/A Res.	<5 bact	5-30 bact	> 30 bact
4	0	1	2	1	-	4	
4	1,5	3	1		3	1	
12	3	4	8		5	5	2
5	4	2	2	1	3		2
8	5 tot 6	8				6	2
5	8	3	2		2	2	1
6	9	3	2	1		6	
5	10 tot 11	1	2	2	2	1	2
5	12	3	2			3	2
8	13	6	2		3	5	
5	14	3	1	1		3	2
4	17 tot 18	2	2				4
3	22 tot 23	2	1		1		2

G/G or G/A = susceptible

A/A = resistant

Coddens et al., 2007. Vet. Microbiol.

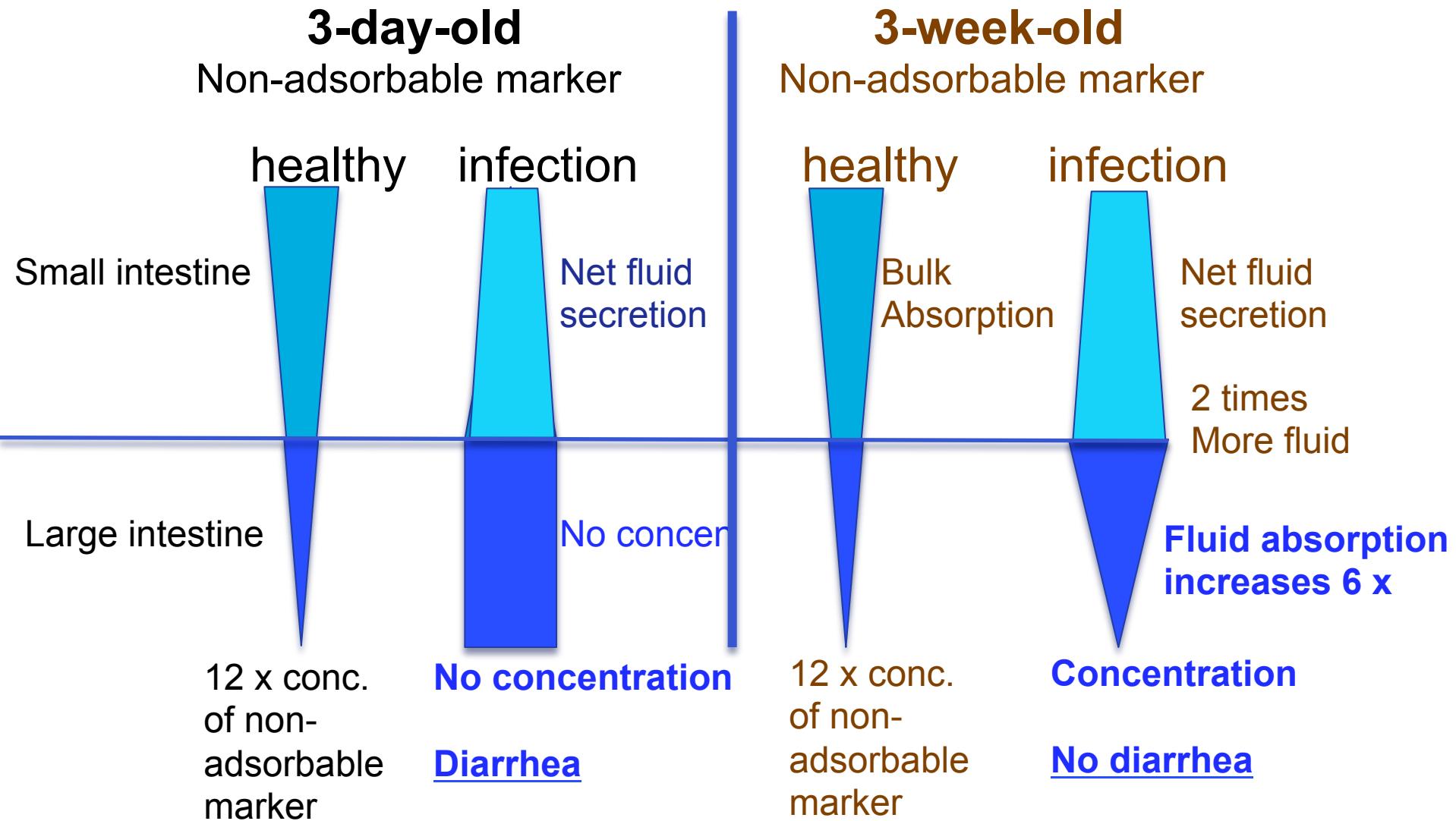


Conclusion

- Expression of the F4R is irrespective of age
- Expression of the F18R is age-dependent
- There is sufficient F18R expression from 3 weeks of age in F18R positive pigs

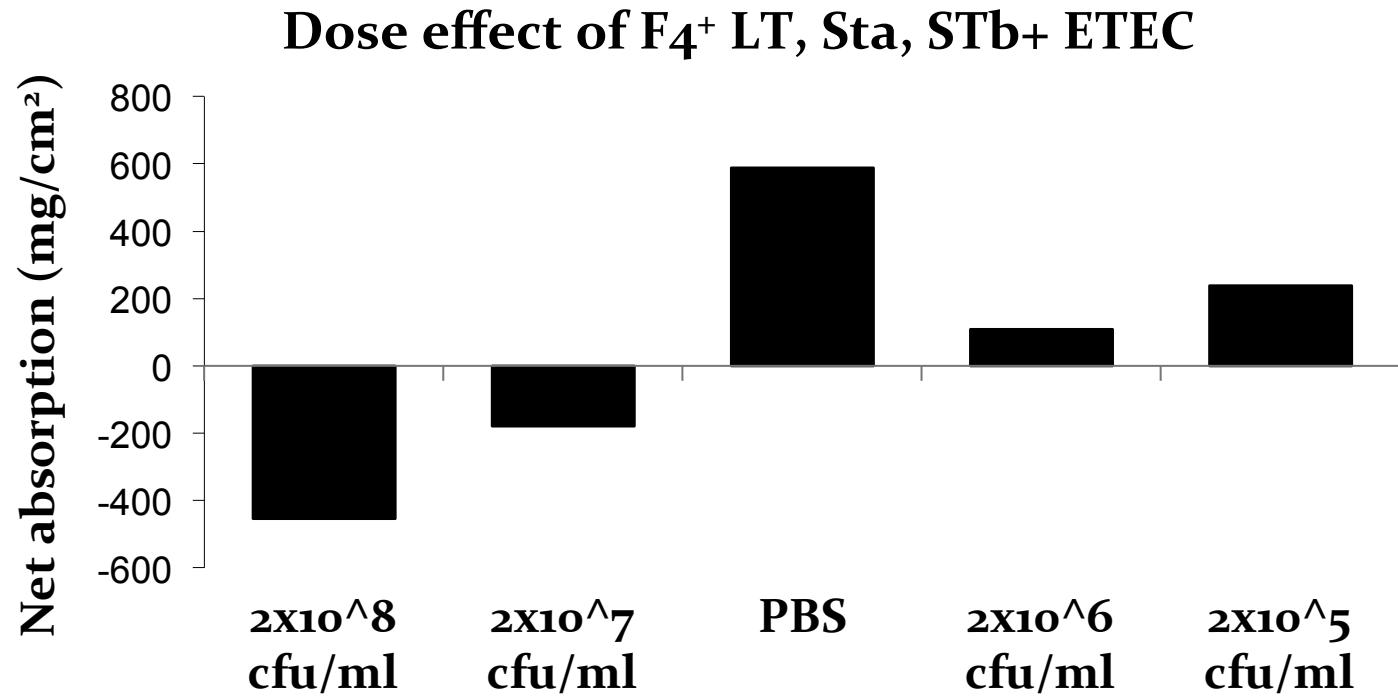
Enterotoxins induce diarrhoea

The weaned pig's large intestine has a higher capacity to absorb fluid

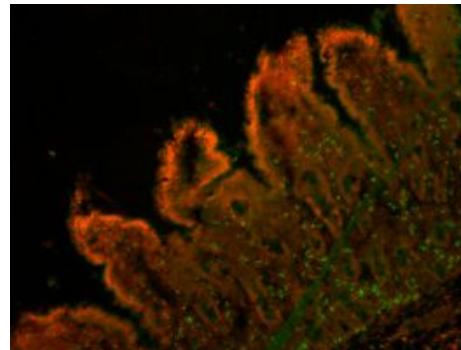


SMALL INTESTINAL SEGMENT PERfusion (SISP)

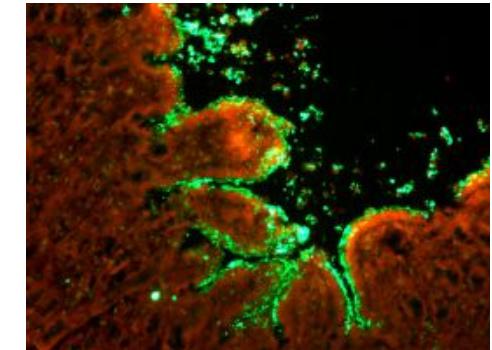
F4R⁺ pig
8h perfusion



Adhesion of the F4⁺bacteria:



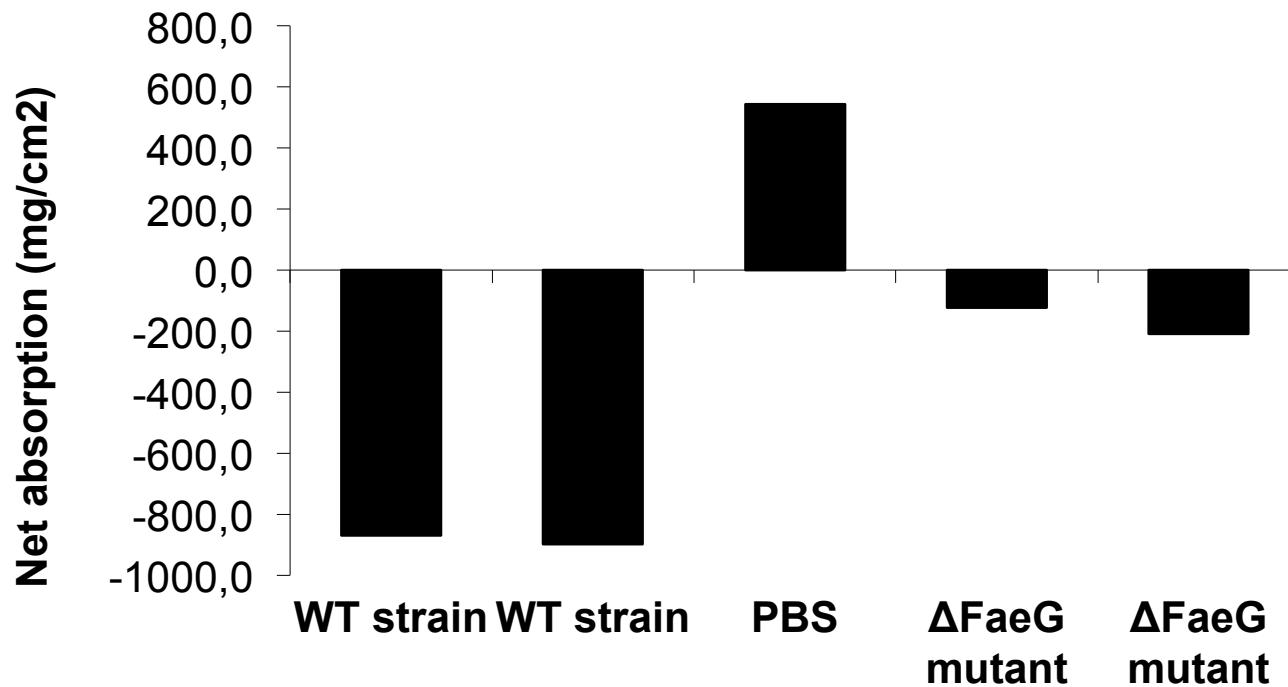
PBS



ETEC GIS26

IMPORTANCE OF F4 MEDIATED ADHESION

- F4R⁺ pig, 8h perfusion
- Compare wild type with mutant strain lacking F4 (deletion of FaeG subunit)



F4 mediated adhesion not necessary but stronger effect

Deletion mutants: phenotypes

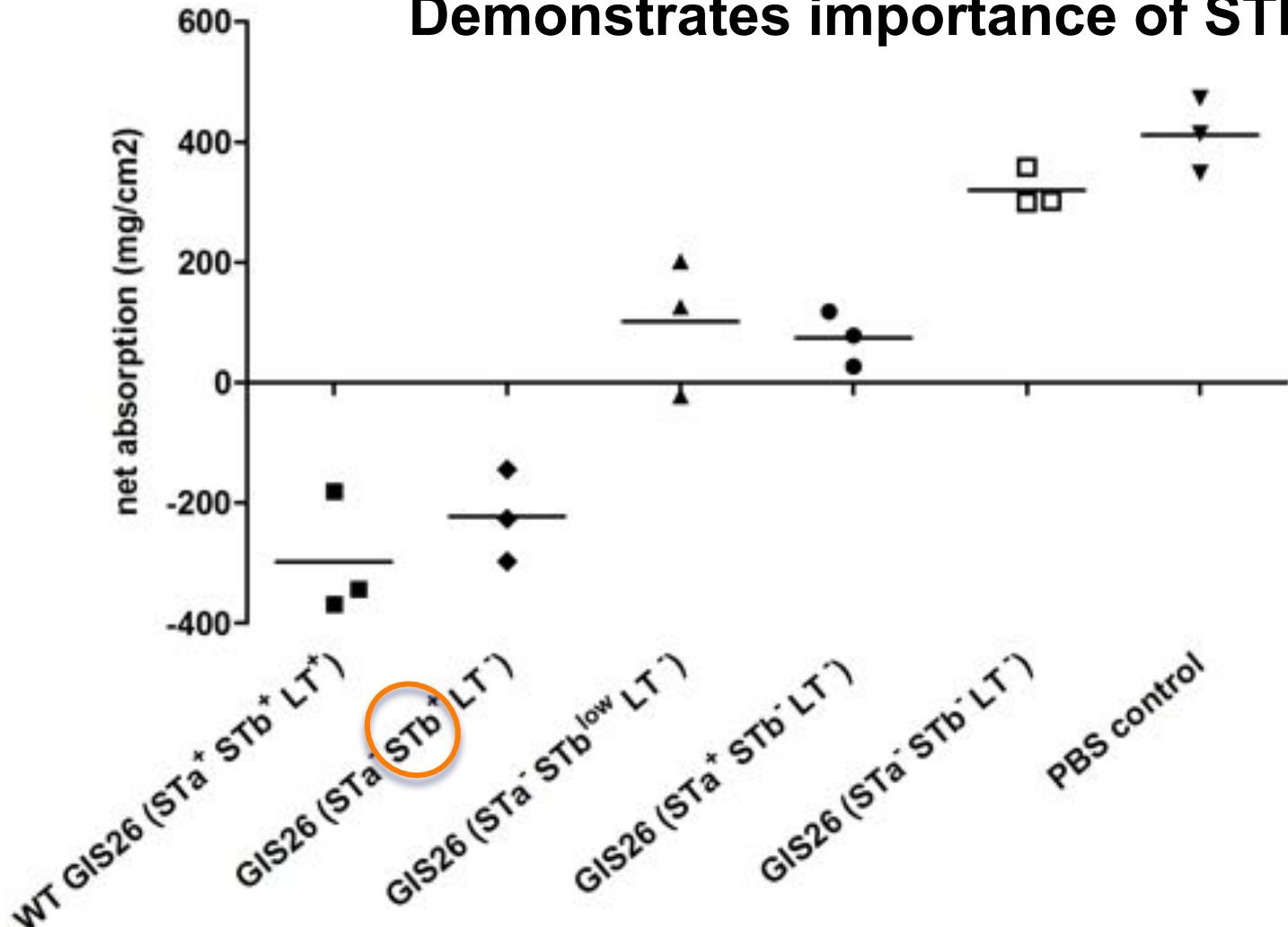
- Quantitative detection of enterotoxin expression *in vitro*
- Detection limits in ng range
- Some mismatches with genotype: gene regulation?

Genotype	Phenotype			Strain designation
	STa	STb	LT	
wild type	+	+	+	GIS26(STa ⁺ STb ⁺ LT ⁺)
ΔeltAB	-	+	-	GIS26(STa ⁻ STb ⁺ LT ⁻)
ΔestBΔeltAB	+	-	-	GIS26(STa ⁺ STb ⁻ LT ⁻)
ΔestA	-	Low	-	GIS26(STa ⁻ STb ^{low} LT ⁻)
ΔestAΔestB:KAN	-	-	-	GIS26(STa ⁻ STb ⁻ LT ⁻)

Small intestinal segments were infected and subsequently perfused during 4 hours

Demonstrates importance of STb

N = 3 F4R⁺

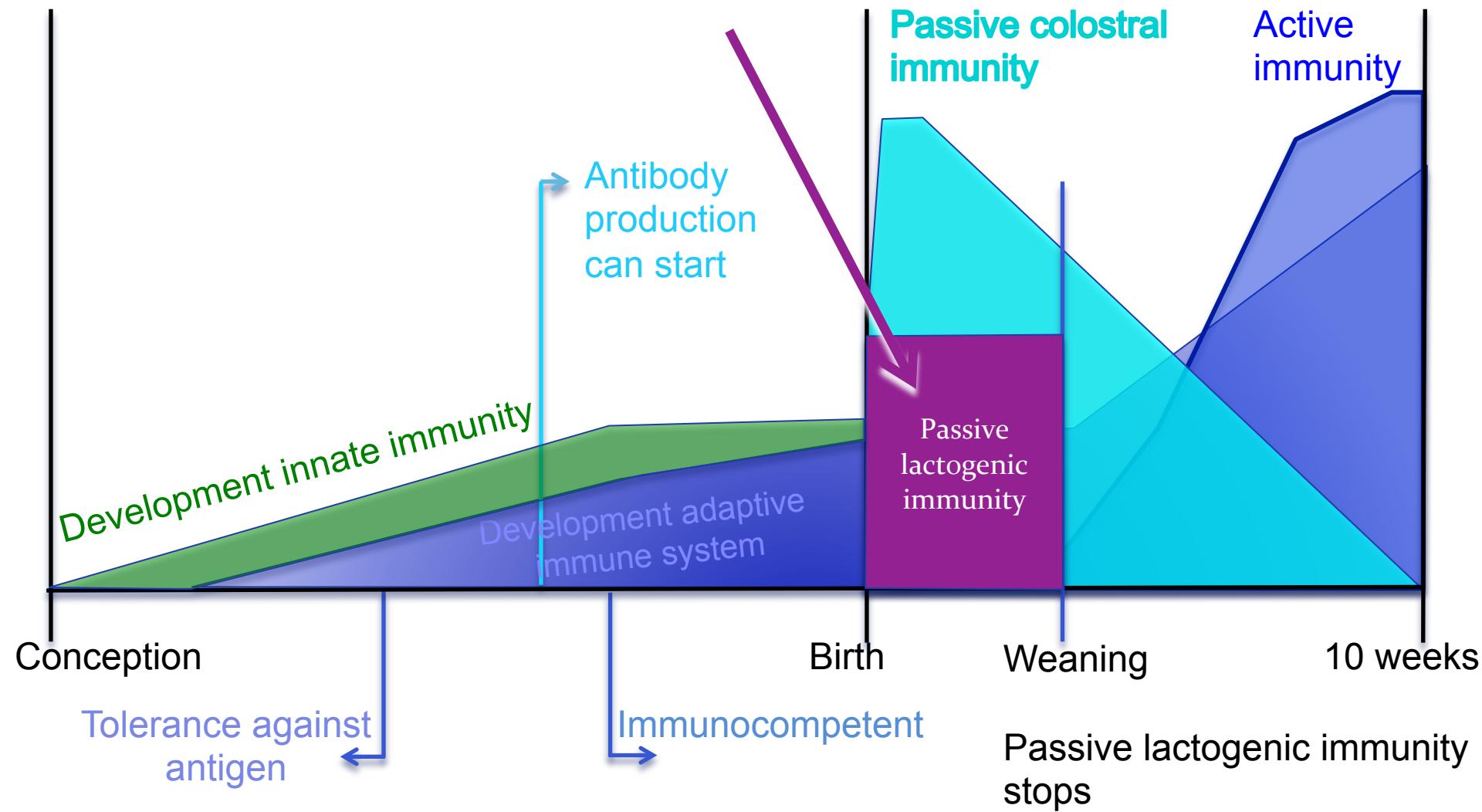


The weaning period

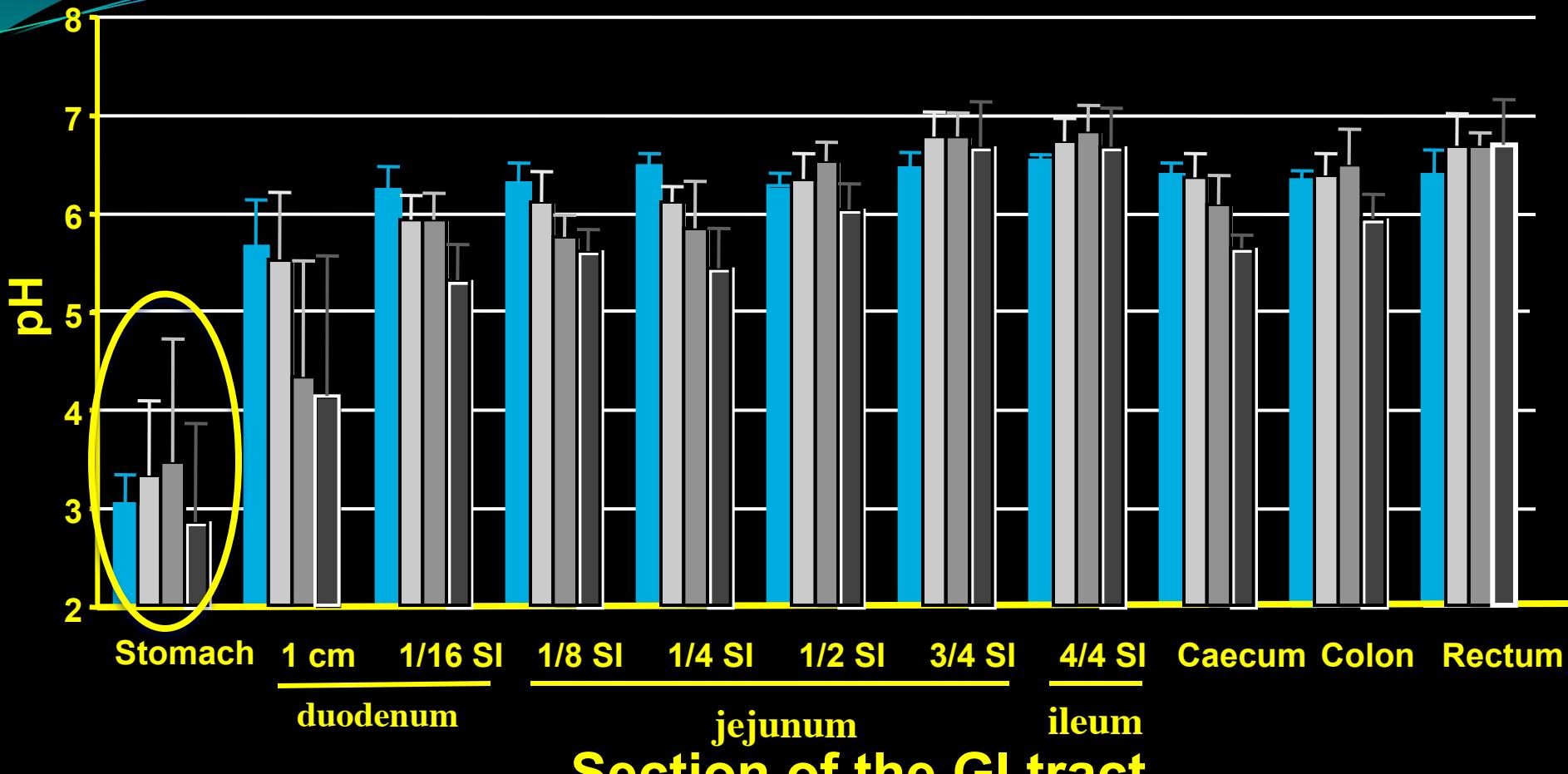
Passive Immunity and mucosal immune response

- Neutralize enteropathogens
- Neutralize oral vaccines/antigens
- Prevent intestinal immune response

Cox, UGent, 2013



pH of gastrointestinal contents



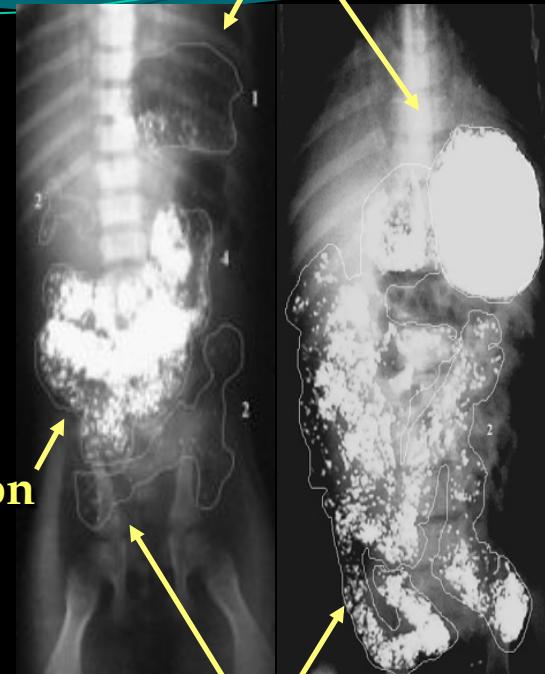
Section of the GI tract

- Suckling piglets n=6
- 3 days postweaning n=6

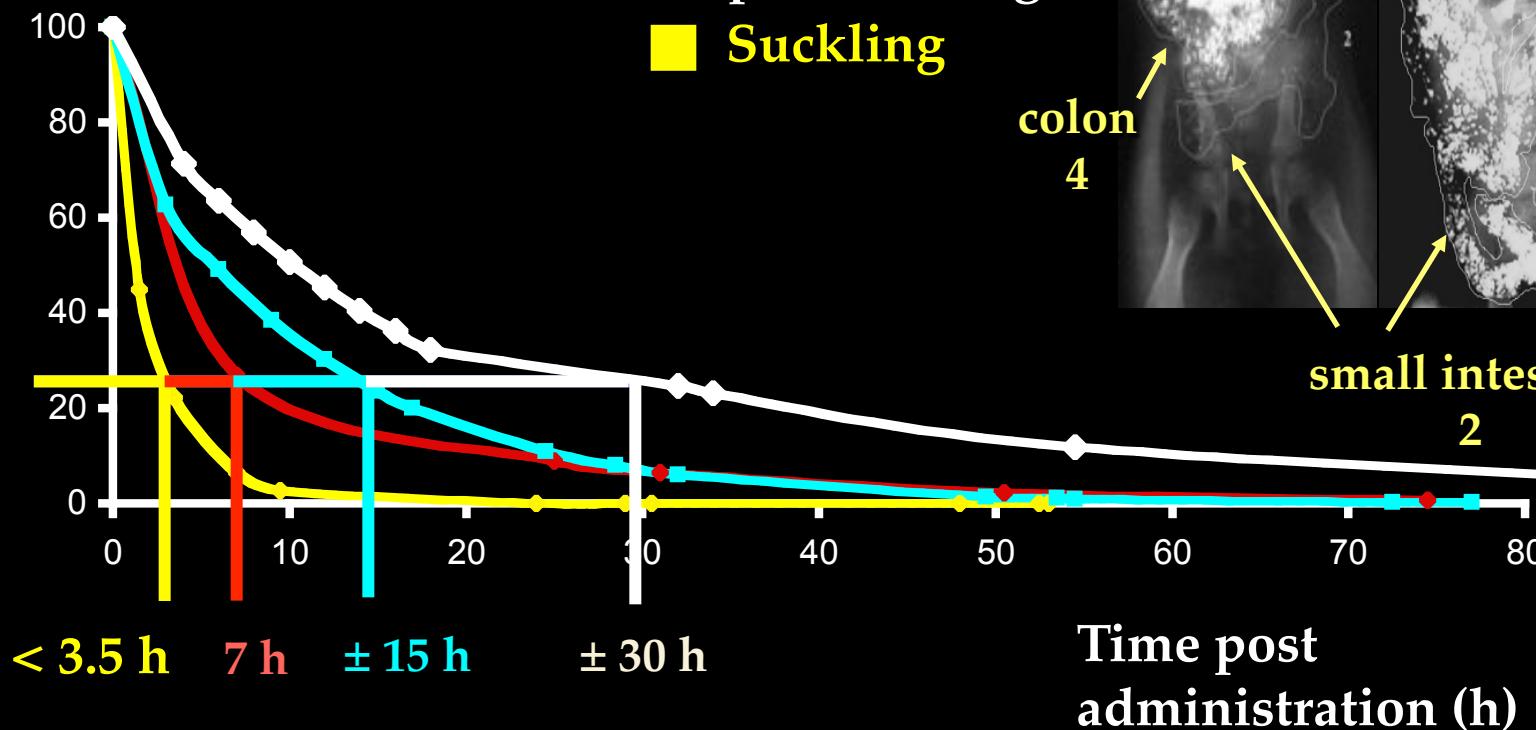
- 1 week postweaning n=6
- 2 weeks postweaning n=6

Gastric emptying

Stomach



% pellets/particles
in the stomach



Gastric emptying for liquids = fast = in suckling piglets 100% within 2 hours

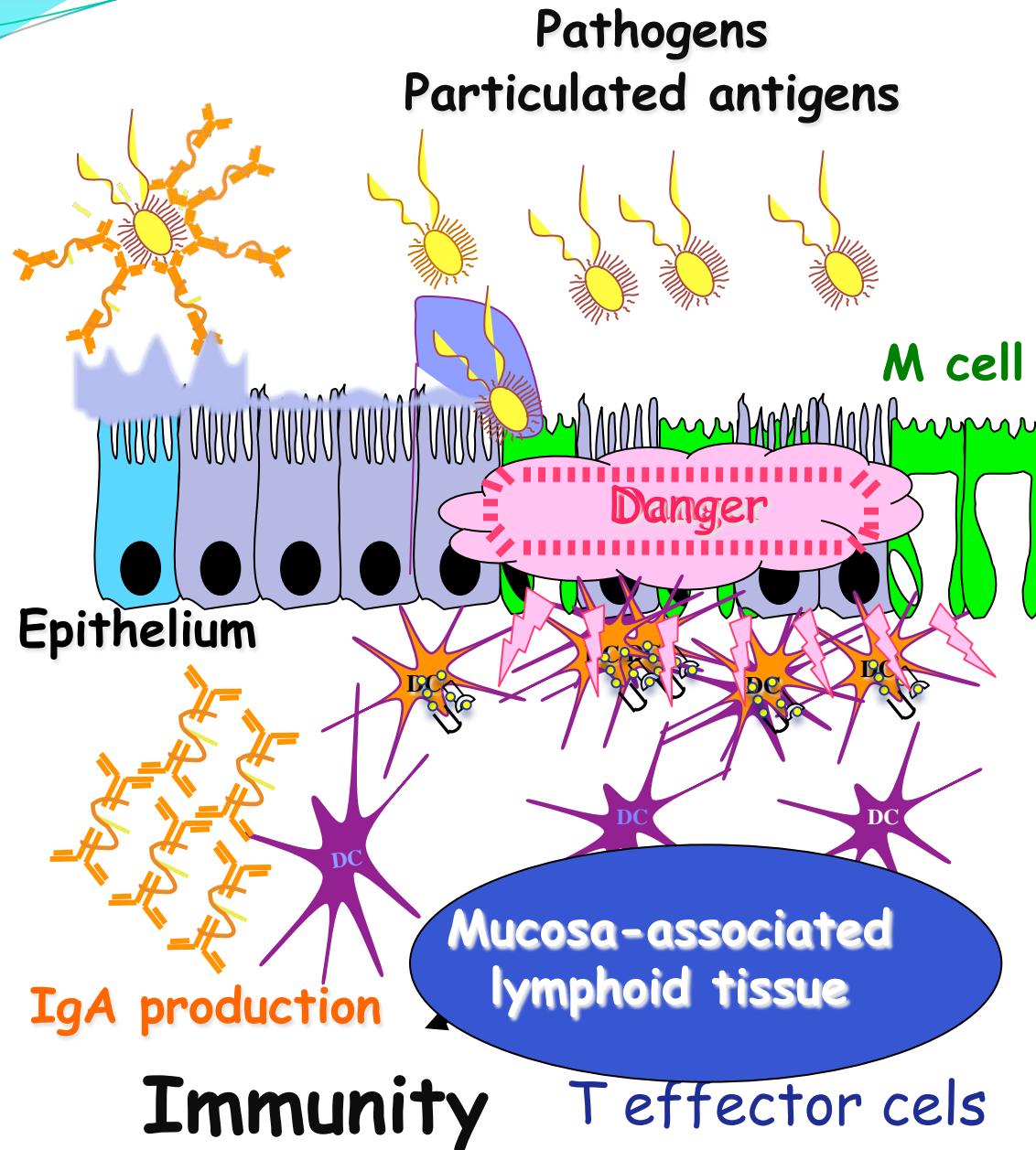
Snoeck et al., 2004. Journal of Controlled Release 94, 143– 153

Changes in Gastrointestinal Physiology

- Proximo-distal gradient reduction in **villus length** along the small intestine
- Decreased activity of most **pancreatic enzymes**
- Decreased activities of **brush border enzymes**
- Increased **para-cellular permeability** in proximal jejunum
- **Trans-cellular permeability** decreased in proximal jejunum and increased in the mid-jejunum
- Activation of **mucosal mast cells, enteric nerves, prostanoid pathways**
- A broad spectrum of **adaptive immune variables suppressed** the first week after weaning
- Several variables of the **innate immune system seem to be stimulated** immediately after weaning
- Transient **over-expression of inflammatory cytokines (IL-1, IL-6, TNF-alfa)** along the intestine

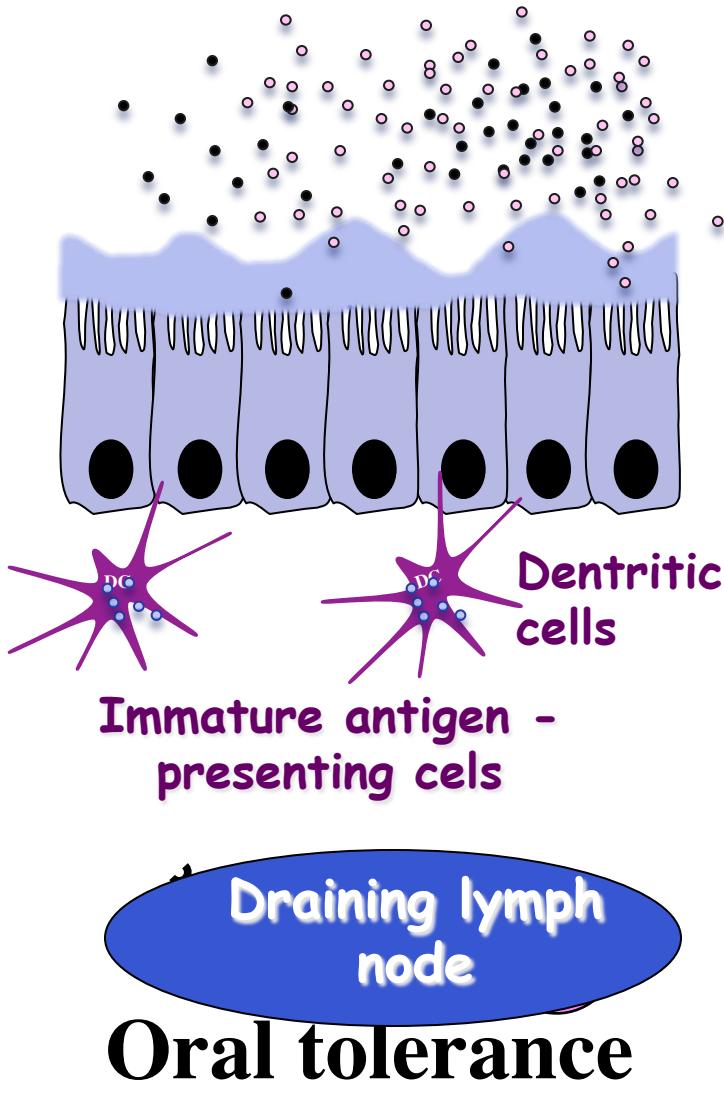
Early and late Immune response

FOLLICLE-ASSOCIATED EPITHELIUM



MUCOSA

Non-replicating soluble antigens



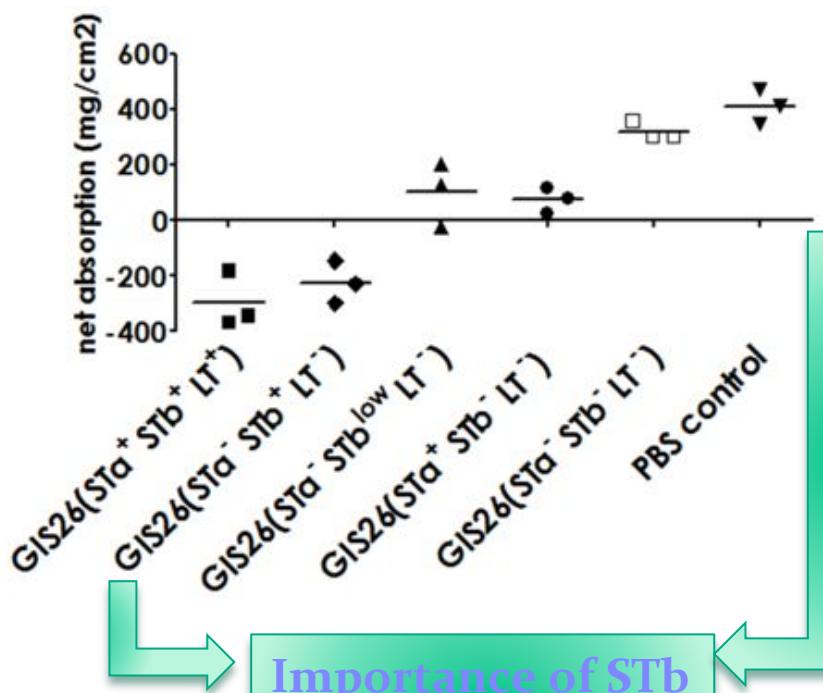
THE EARLY RESPONSE MICRO-ARRAY ANALYSIS



- 3 F4R⁺ pigs different gut segments infected with
 - **Uninfected control versus wild type** strain infected segment = what is the general ETEC response:
 - **Mutant strain versus wild type** strain infected segment = What is the role of enterotoxins:
- mRNA from intestinal segments after 4h perfusion
- Porcine Genome Array (Affymetrix 23,937 probe sets → 20,201 *Sus scrofa* genes)



Functional response



Gene expression

13 transcripts up-regulated by GIS26 wild type

Gene	Gene symbol	Probe Set ID	Log2 ratio
Matrix metalloproteinase 3	MMP3	Ssc.15927.1.S1_at	4.18
Interleukin-17A	IL17A	Ssc.15927.2.S1_at	4.16
Pancreatitis associated protein	PAP (REG3A)	Ssc.15927.2.A1_at	2.90
Interleukin-1 beta	IL1B	Ssc.16470.1.S1_a_at	3.68
Interleukin-1 alpha	IL1A	Ssc.17573.1.S1_at	3.10
Dual oxidase 2	DUOX2	Ssc.15601.1.A1_s_at	2.72
Interleukin-1 receptor antagonist	IL1RN	Ssc.33.1.S1_at	2.65
Matrix metalloproteinase 1	MMP1	Ssc.30857.1.S1_at	2.68
Ectoderm-neural cortex protein 1	ENC1	Ssc.16013.1.S1_at	2.33
		Ssc.113.1.S2_at	2.05
		Ssc.113.1.S1_at	2.05
		Ssc.16250.1.S2_at	2.01

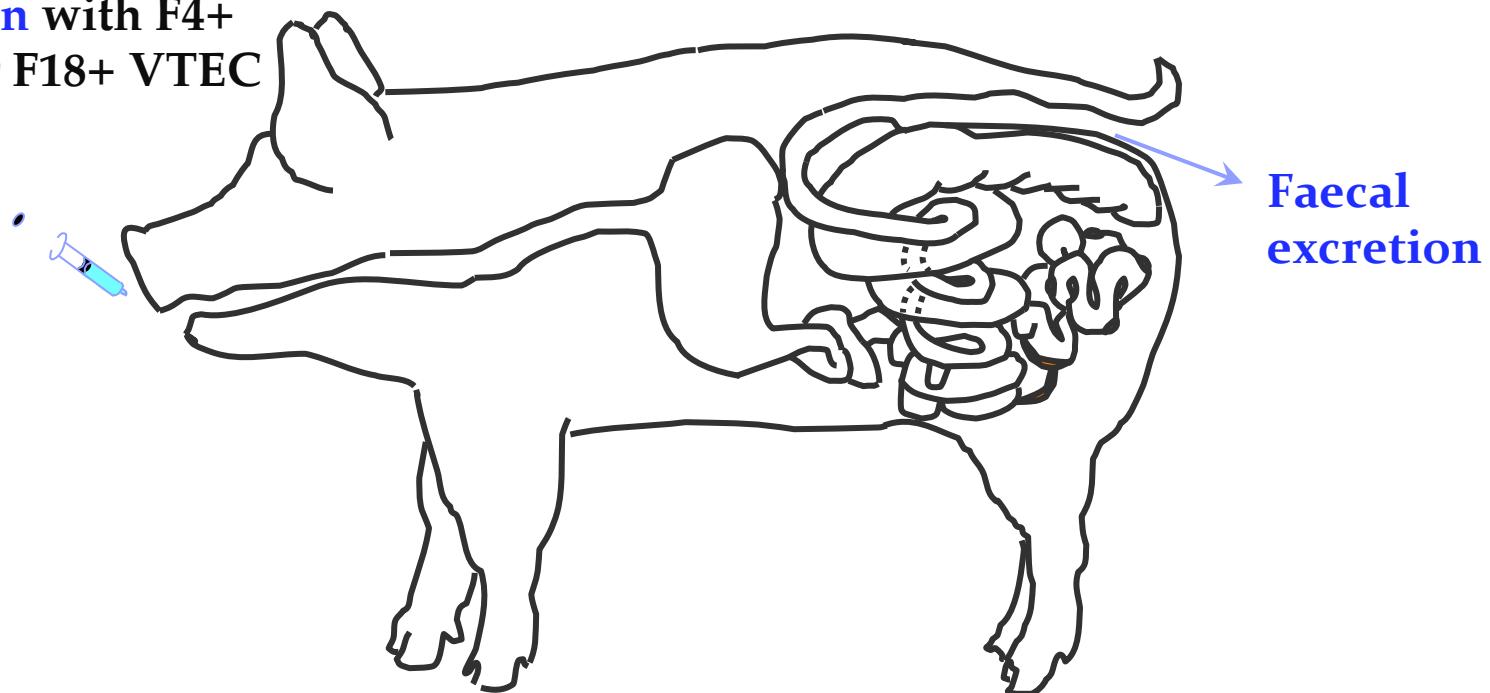
Conclusions:

ETEC induces

- 1) non-toxin related general antibacterial response (PAP, MMP1, IL8,...)
- 2) Important role for STb in small intestinal secretion as well as in the ETEC induced immune response (MMP3, IL17, IL1)

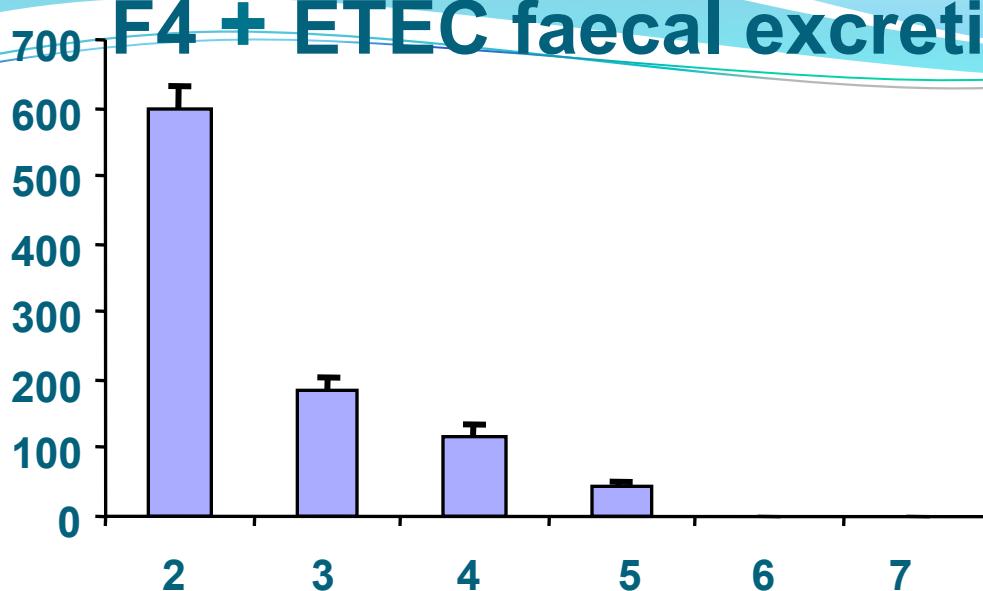
Adaptive Immune response following infection with F4+ ETEC or F18+ VTEC

Oral **infection** with F4+
ETEC or F18+ VTEC



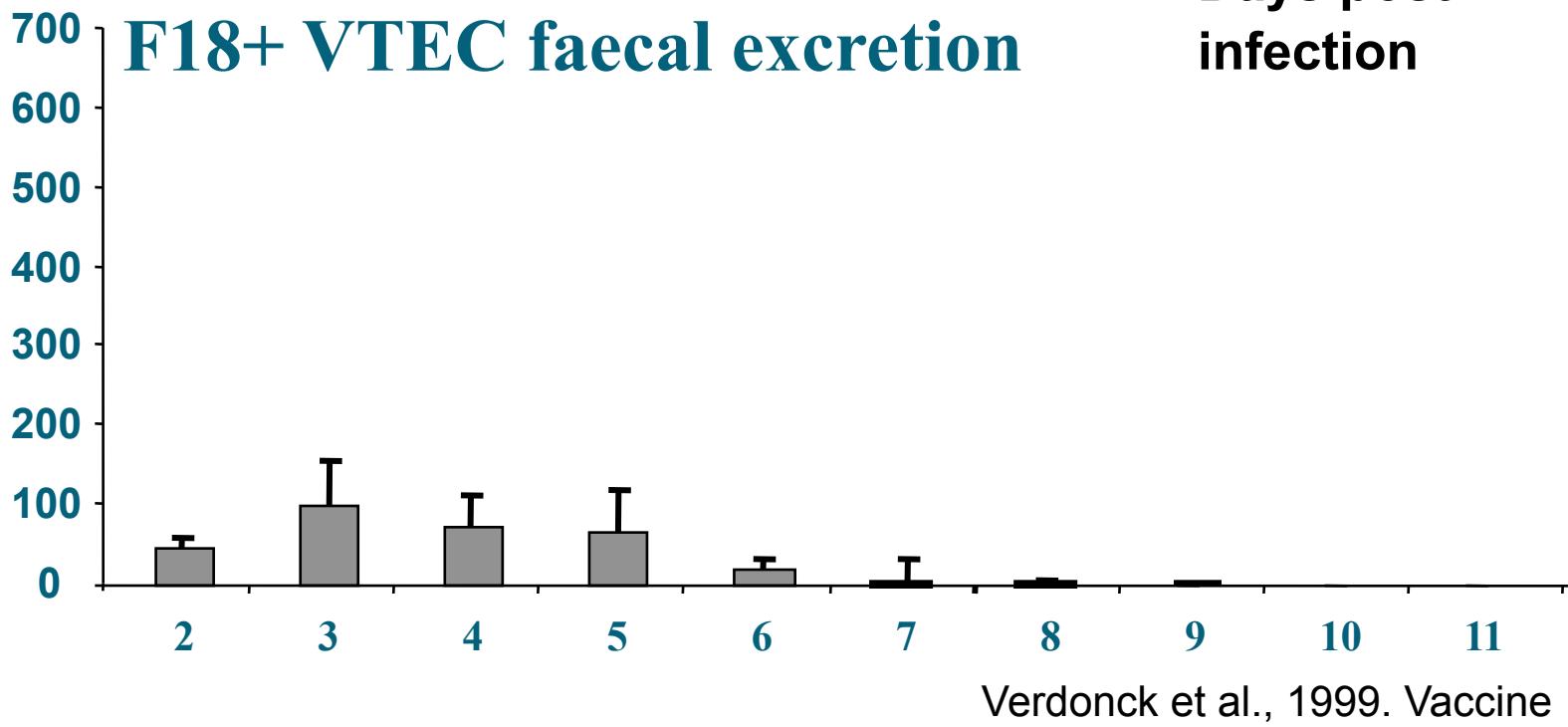
F4 + ETEC faecal excretion

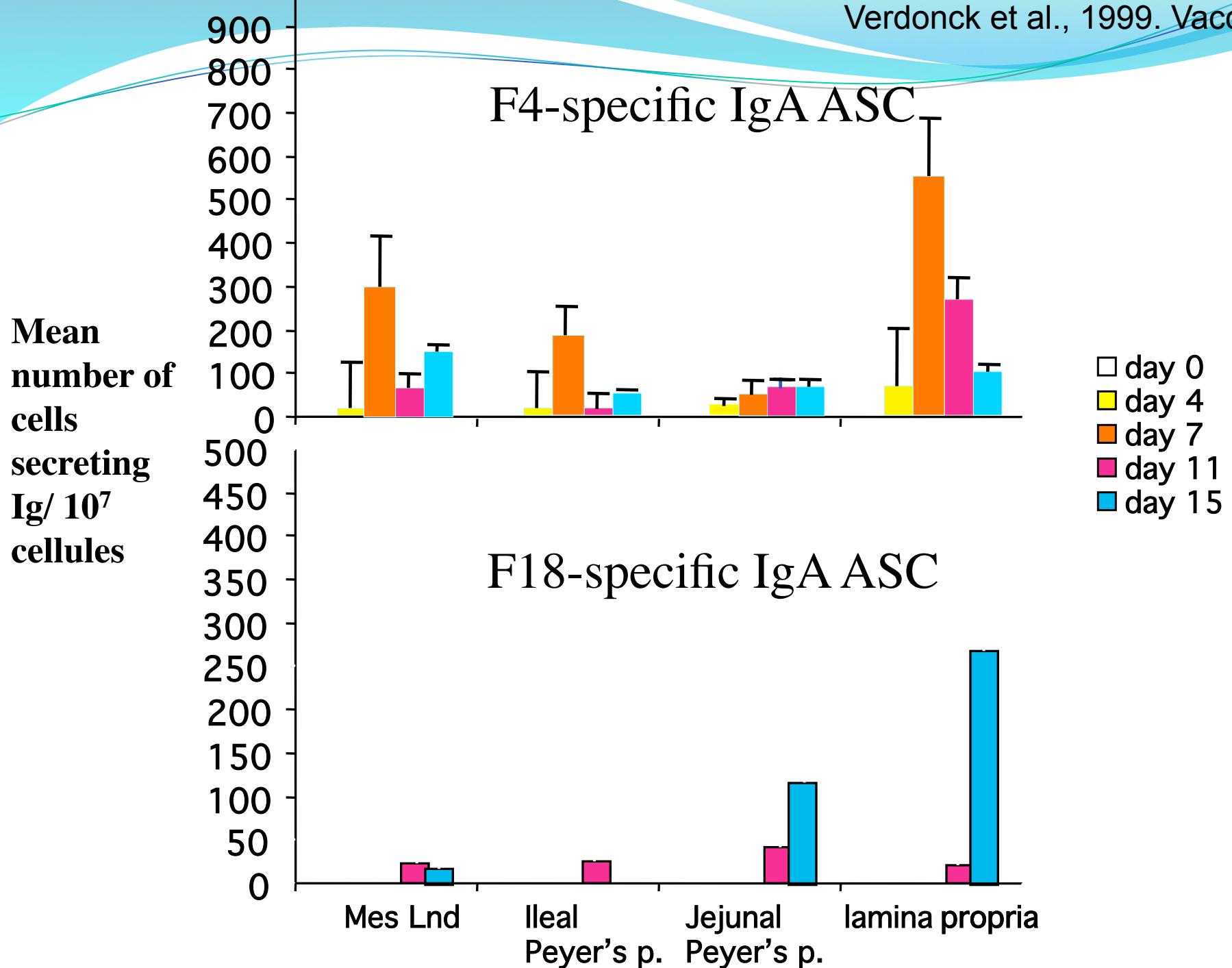
10^6
E. coli
per g
faeces



F18+ VTEC faecal excretion

Days post
infection





Conclusions

Infection with F4+  F18+ *E. coli*

- Colonization difference

- Rapid (1st week)
- High



Slower (2nd week)

Lower excretion

- Difference in antibody response

- Quick



Slow

Different fimbriae (affinity)

Different toxins (enteroxins => secretion; STb => inflammatory, LT => adjuvants)

Immunization with F4 or F18 fimbriae via the oral route

2. Infection challenge with

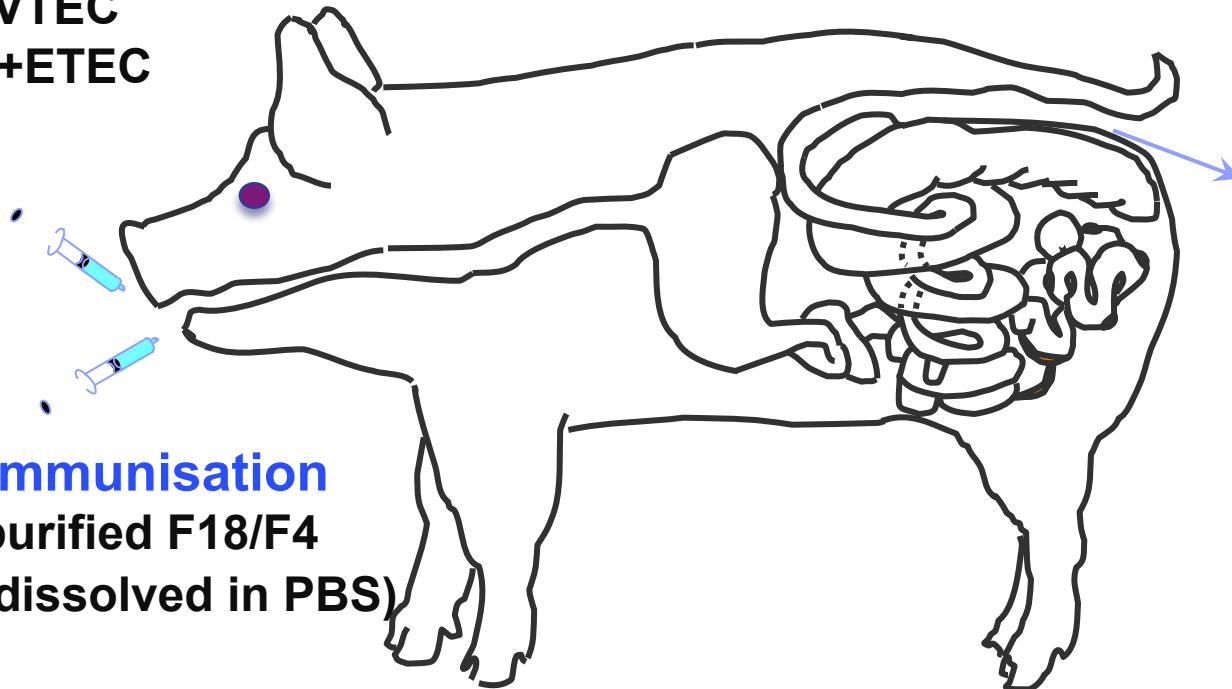
F18+ VTEC

or F4+ETEC

1. Oral Immunisation

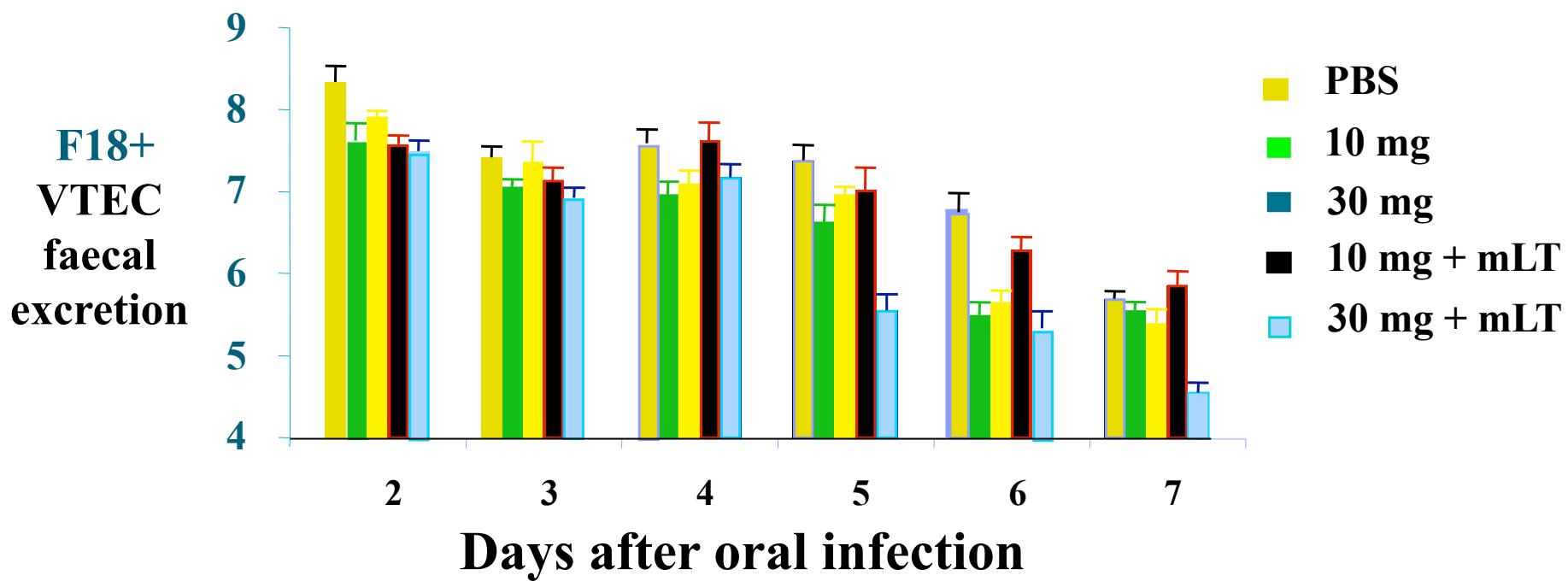
with purified F18/F4

(and dissolved in PBS)

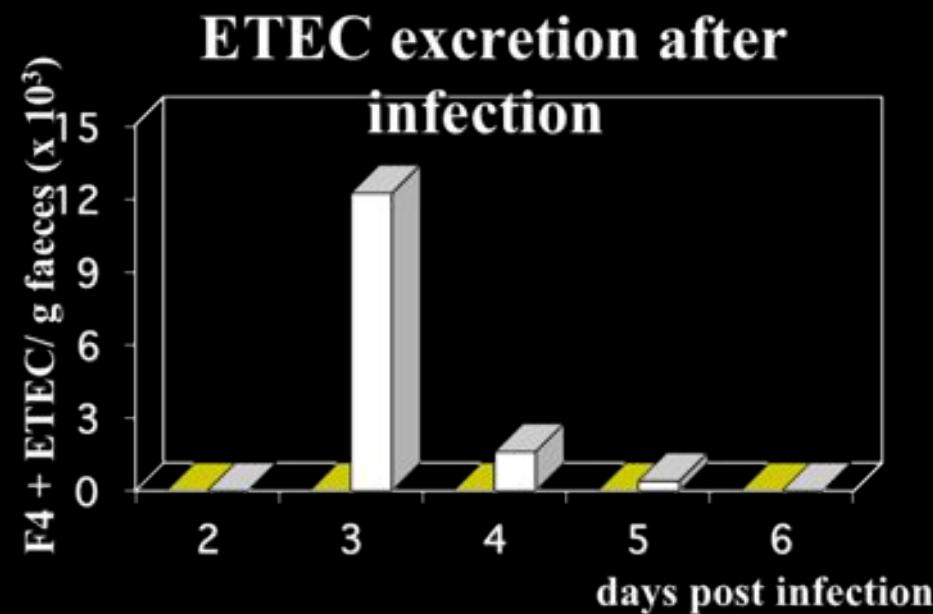
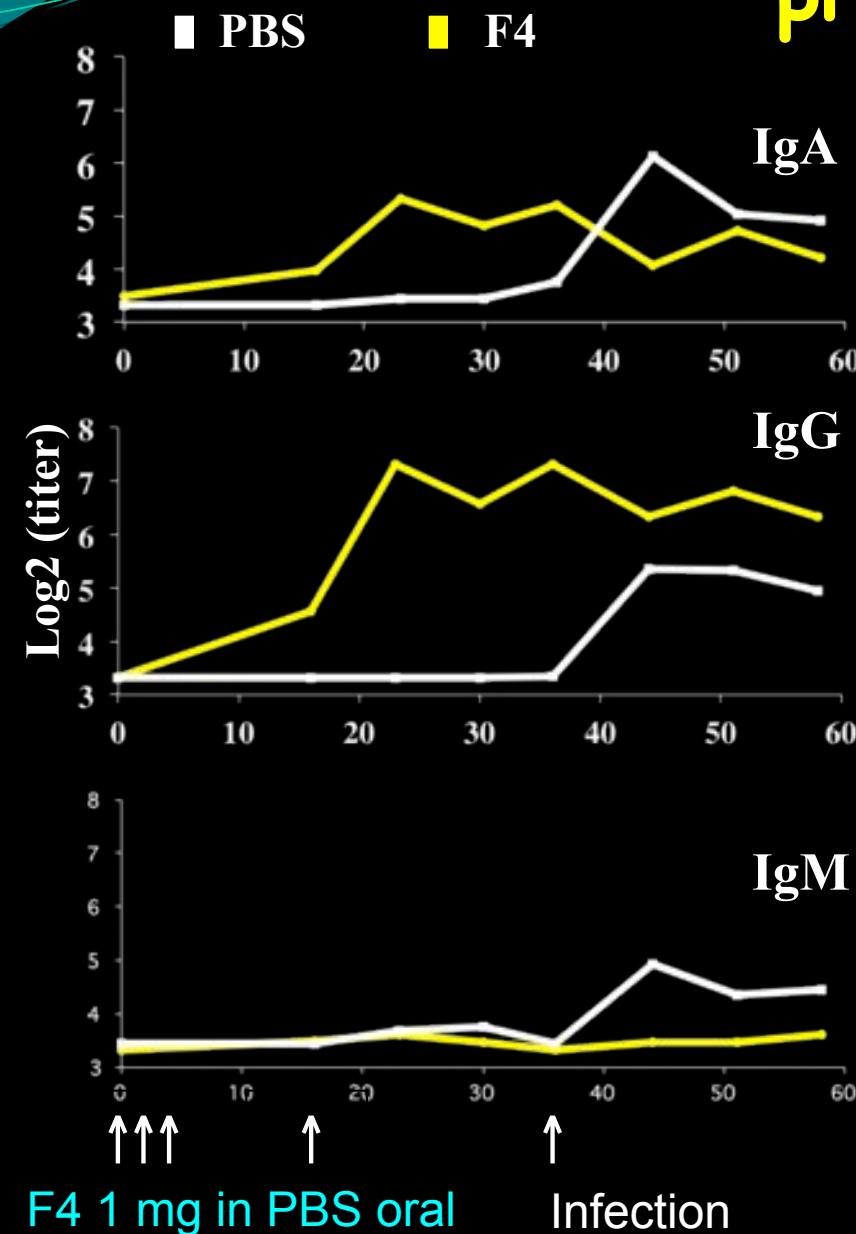


3. Faecal excretion of
F18+ VTEC/
F4+ ETEC

Oral vaccination with F18 fimbriae does not protect



The mucosal response against F4 is protective



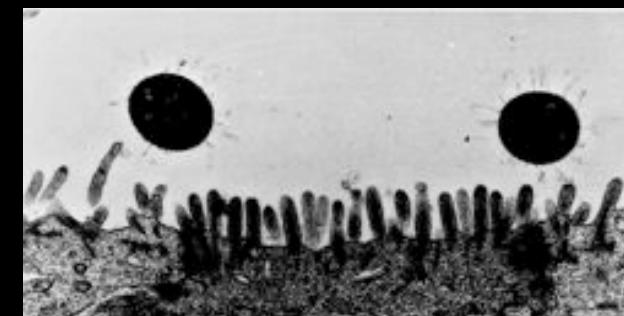
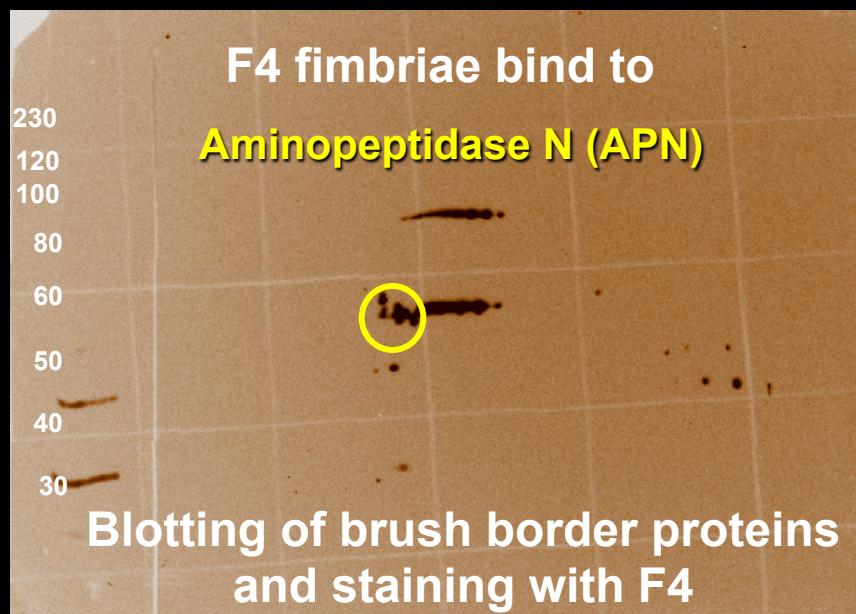
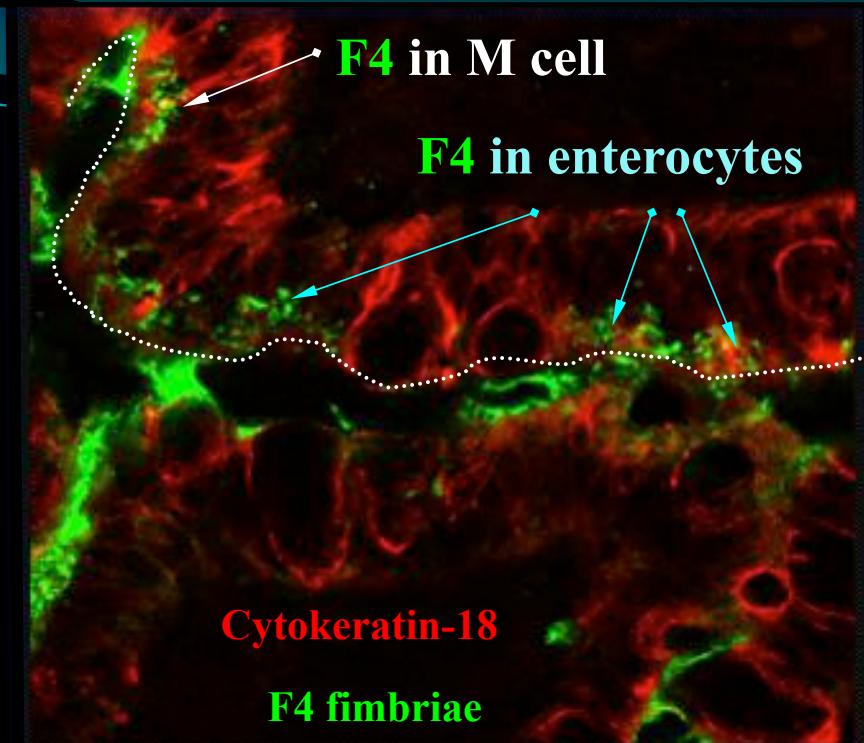
=> Oral F4 induces
protective mucosal
response !

Binding and uptake of F4 fimbriae

Ligated loops injected with F4



Snoeck et al., 2008. Vet Imm Immunopath.



Melkebeek et al., 2012. Mucosal Immunology

Summary

- Weaning = opportunity for ETEC/VTEC to colonize the intestine
 - Decreased food intake
 - Decreased gastrointestinal transit
 - Decreased digestion
 - Absence of passive protection
 - Suppression of adaptive immunity
- Virulence factors influences colonization and immunity
 - Different fimbriae => different affinity => slower or faster adhesion
 - Different fimbriae differ in immunogenicity
 - Enterotoxins induce fluid secretion influencing colonization
 - Enterotoxins can influence the immune response => inflammation, adjuvanticity
- Immunity should neutralize toxins and/or at least prevent colonization in the very early phase

Acknowledgements

Former PhD students)

Prof. Dr. Wim Van den Broeck

dr. Frank Verdonck (Ablynx)

dr. Petra Tiels (VIB-UGent)

Karien van Gog (practice)

dr. Veerle Snoeck (Ablynx)

dr. Kristien Rasschaert (B&D)

dr. Annelies Coddens (Ablynx)

Lic. Philippe Bellot (AsA4)

Lab of Immunology (UGent)

Prof dr. Bruno Goddeeris (Lab Immuno/KULeuven)

dr. Vesna Melkebeek

dr. Bert Devriendt

Vrije Universiteit Brussel (VUB)

Prof. Dr. Henri Degreve and Han Remaut

Lab of Pharmaceutical Biotechnology (Ugent)

Prof dr. Dieter Deforce and dr. Kelly Tillemans

Department of Biosystems (KULeuven)

Prof dr. Theo Niewold and dr. Marisa Geens



Faculty of Veterinary
Medicine



Cox, UGent, 2013



University of Gothenburg, Sweden

Prof. dr Susann Teneberg

Financial support



Fonds Wetenschappelijk Onderzoek
Research Foundation - Flanders